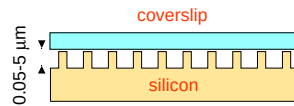


Microfabricated arrays

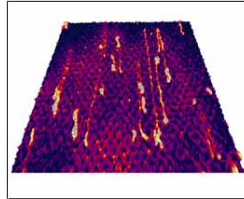
Volkmut & Austin '92

2-dimensional obstacle course for DNA



Advantages:

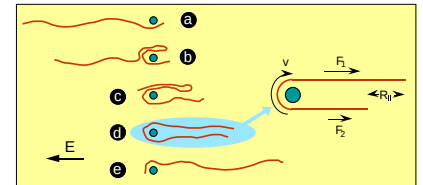
- well-defined & **regular**
- can **observe** DNA dynamics
- can **choose** design



Collisions with posts

Volkmut et al. '94

Episodic motion in a regular lattice



- typical engagement time $\sim L$,
... but typical collision frequency $\sim 1/L$

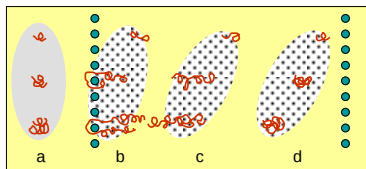


fails to provide effective separation

Separation by collisions with posts

Duke et al. '97

Rows of posts



- collision **hinders** molecule
- delay depends on length L

$$t_{disengage} \sim \frac{L}{2\mu_0 E} \log \left(\frac{L}{R_{||}(0)} \right)$$

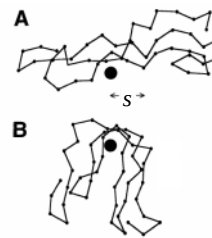
Require DNA to relax in the time taken to travel own length

- works best for small DNA molecules
- could sequence 500 nucleotides in 10 mins, using 20 nm posts, separated by 400 nm

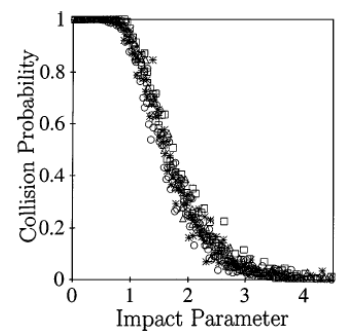
Collisions between DNA and a point obstacle

Sevick & Williams '96

Strong-stretching regime

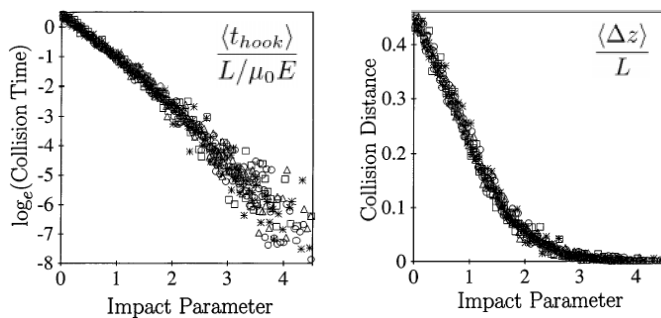


$$\text{Impact parameter } \frac{s}{\langle R_x \rangle}$$



Collisions between DNA and a point obstacle

Sevick & Williams '96

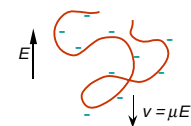


Electrophoretic stretching

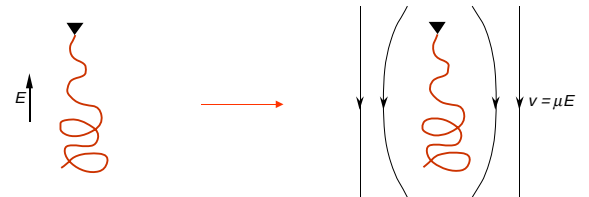
Long, Viovy & Ajdari '96

Electrophoresis:

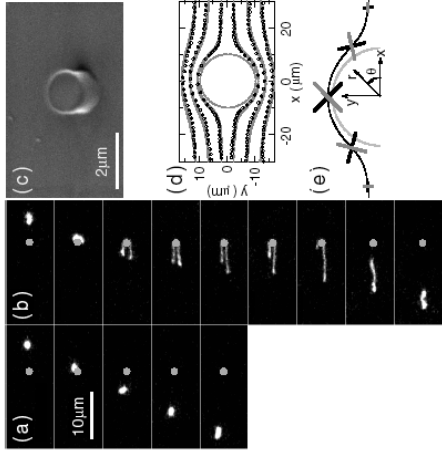
- hydrodynamic interaction **screened**
→ μ independent of size



DNA tethered in a field:



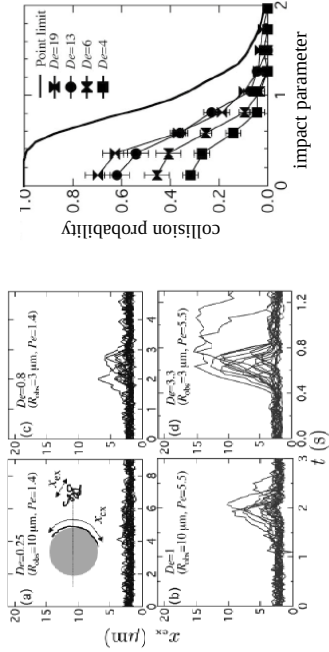
- hydrodynamics **not screened** ... equivalent to DNA tethered in a flow



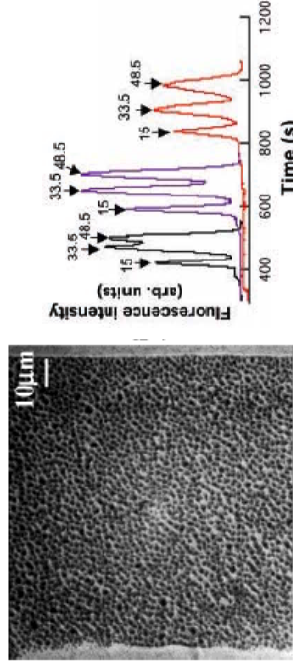
DNA-post collisions

DNA-post collisions

Owing to extensional flow, collision probability depends also on a dynamical parameter $De = \frac{2\mu E}{R_{obs}} \tau_{relax}$



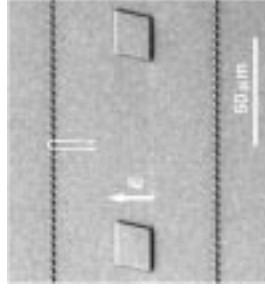
Separation in self-assembled magnetic matrices



Electrophoretic stretching of DNA in a confined space

DNA confined in a **thin slit**, depth **h**

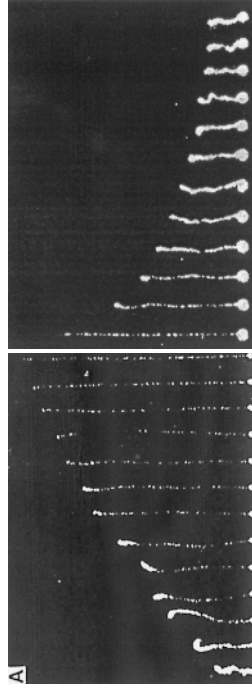
- driven by electric field
- transient stretching & relaxation



Hydrodynamic interactions **screened** on scale $> h$

Prediction: **drag increases as depth decreases**

Extension of tethered DNA in a flow



Stem & flower model

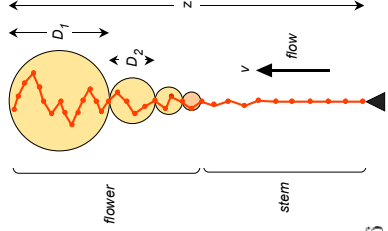
Polymer appears as a stretched **stem**, capped by a **flower** of Pincus blobs, opaque to the flow

- Generalize model to include:
- **worm-like chain elasticity**
 - **hydrodynamic interactions** between blobs

blob α contains n_α beads $D_\alpha = \sqrt{\frac{n_\alpha}{3}} b$

effective friction of a bead $\zeta = \frac{\zeta_b}{\sqrt{n_\alpha} \log(z/D_1)}$

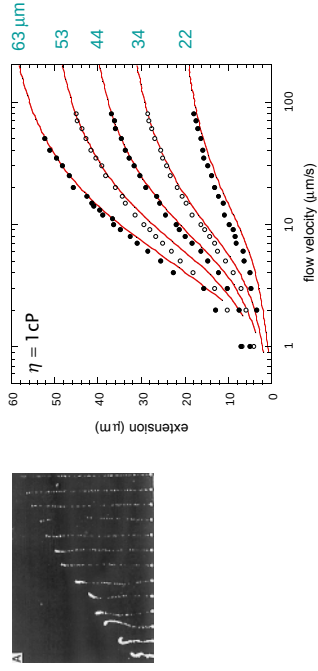
$\zeta_b = A\eta b$ $A \approx 6$



Model equilibrium extension

Perkins et al. '95

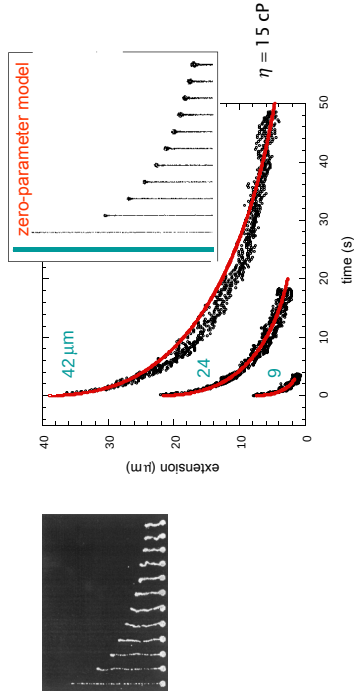
Extension of a DNA molecule tethered in a uniform flow



Model relaxation

Perkins et al. '94

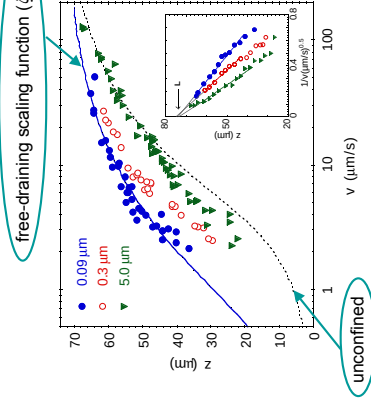
Relaxation of tethered DNA



Thin slit extension vs electrophoretic drift velocity

Bakajin et al. '98

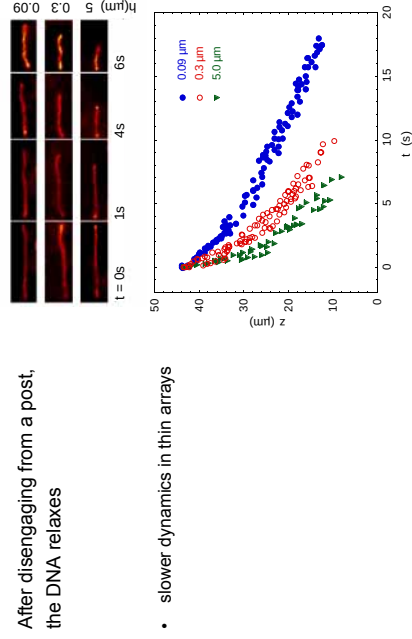
free-draining scaling function ($\zeta \sim L$)



unconfined

Thin slit relaxation dynamics

After disengaging from a post, the DNA relaxes

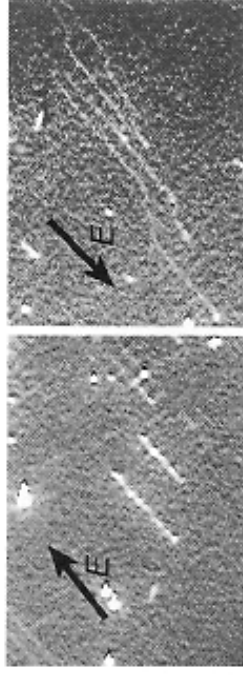


• slower dynamics in thin arrays

Linear migration

Regular array of large posts with well-aligned field

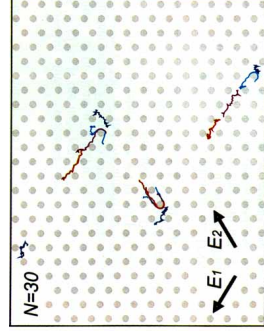
→ DNA migrates straight along channels



Pulsed-field electrophoresis in regular arrays

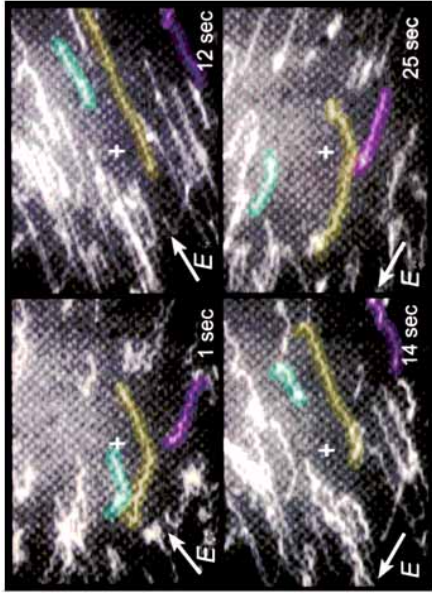
Duke et al. '96

Switch field alternately along two different directions



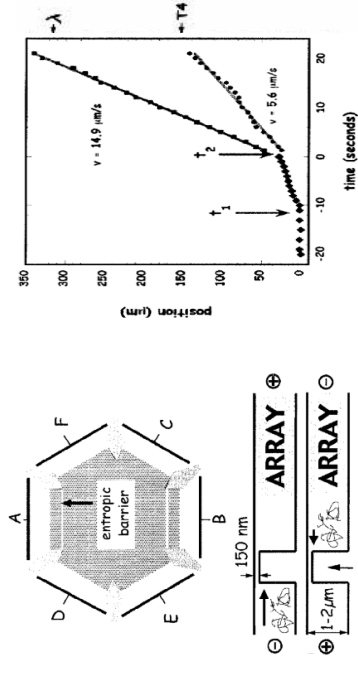
Pulsed-field arrays molecular motion

Duke et al. '96; Bakajin et al. '01



Pulsed-field hex arrays ultra-fast separation

Bakajin et al. '01



Separation of 100 kb DNA in 10 seconds!



Pulsed-field arrays separation

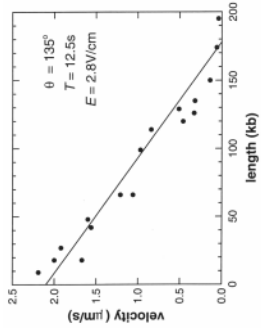
Duke et al. '96

Advantages of arrays:

- regular motion → good resolution
 - high fields → fast separation
 - linear fractionation
- straightforward recipe for choosing pulse parameters

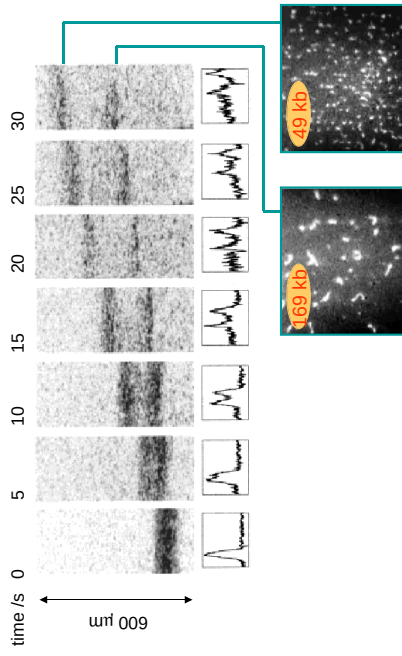
$$v = v_0 \cos(\Theta/2) \left(1 - \frac{L_s}{L_{max}}\right)$$

$$L_{max} = \left(\frac{T}{T_0}\right) \left(\frac{E}{E_0}\right) b$$

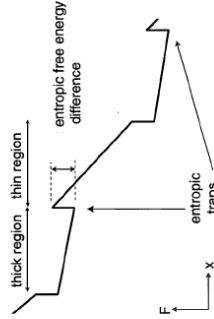
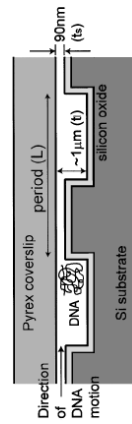


Pulsed-field hex arrays ultra-fast separation

Bakajin et al. '01

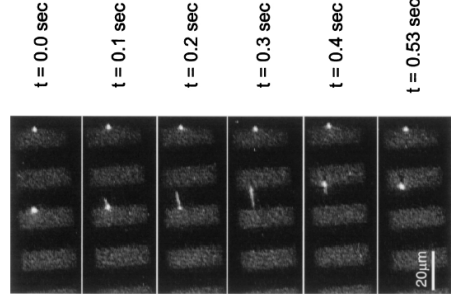


Han, Turner & Craighead '99



Entropic trapping

Han, Turner & Craighead '99

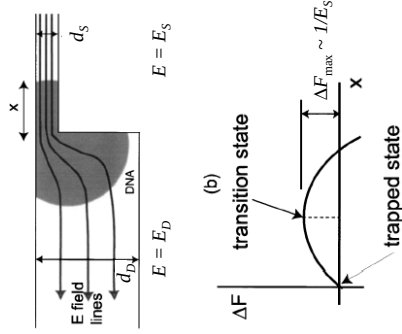


Entropic trapping mobility

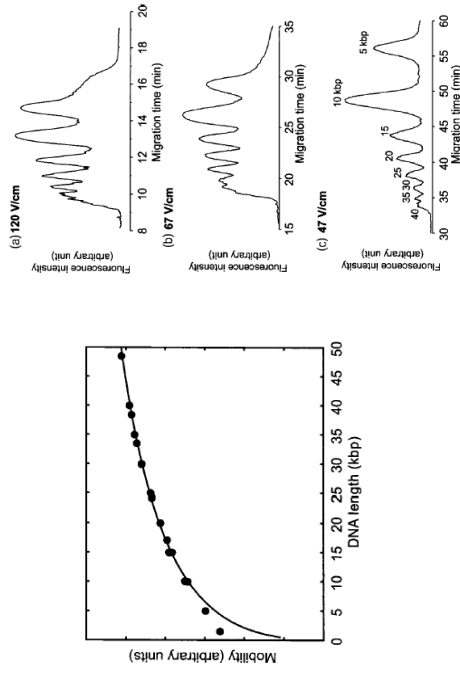
$$\frac{\mu}{\mu_0} = \frac{t_{travel}}{t_{travel} + t_{trap}}$$

$$t_{travel} = \frac{L}{2} \left(\frac{1}{\mu_0 E_S} + \frac{1}{\mu_0 E_D} \right)$$

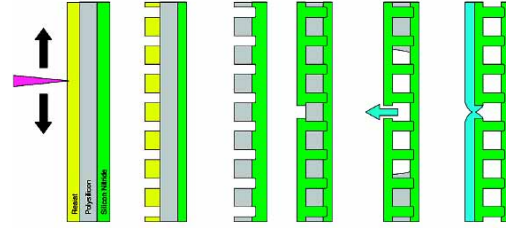
$$t_{trap} \sim \frac{1}{d_S N^{1/2}} \exp \left(\frac{\alpha}{E_S kT} \right)$$



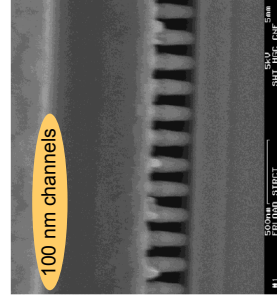
Entropic trapping separation



e-beam lithography



'lost wax' process

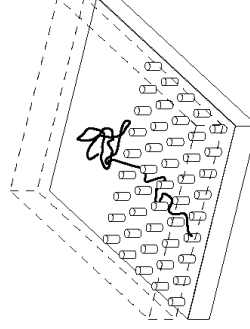


Cabodi, Turner & Craighead '02

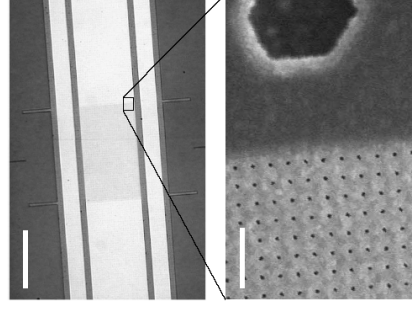
Dense array of posts

DNA is entropically inhibited from entering
forest of posts...

... but can be driven in by a high field

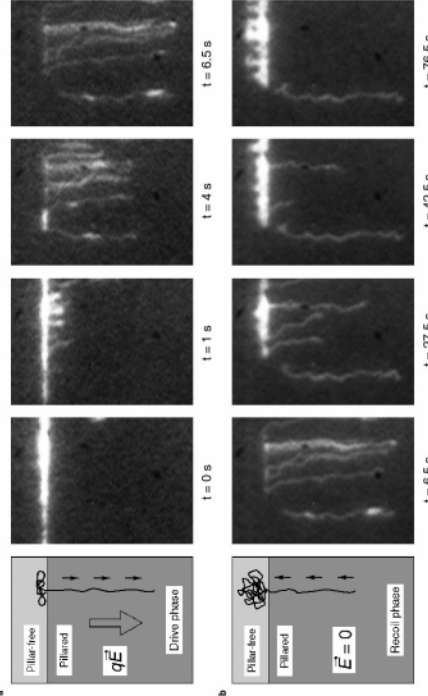


Turner, Cabodi & Craighead '02



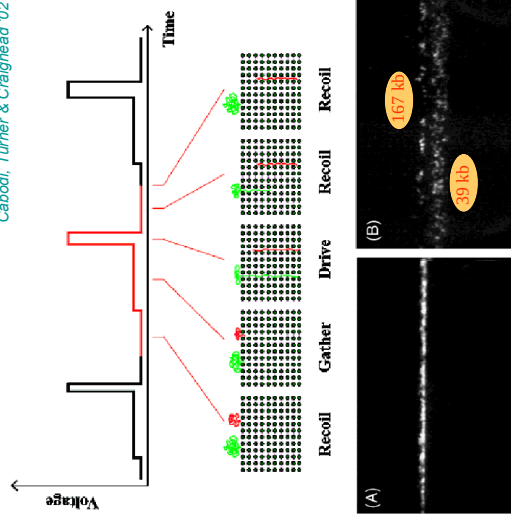
Entropic recoil

Turner, Cabodi & Craighead '02



Cabodi, Turner & Craighead '02

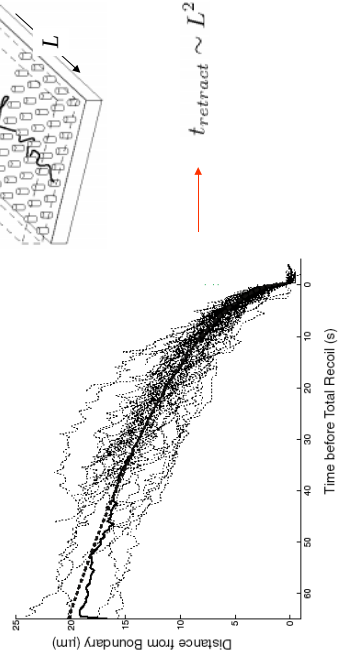
Entropic recoil separation



Entropic recoil retraction time

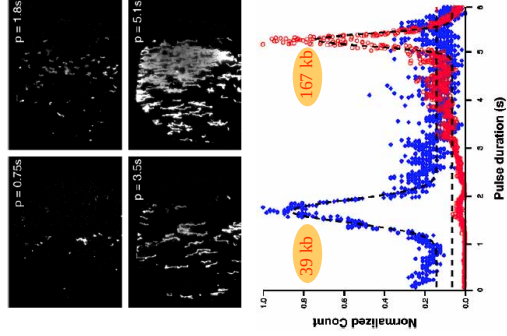
Turner, Cabodi & Craighead '02

- constant retraction force
- friction proportional to unretracted length



Entropic recoil: pulse time ramp

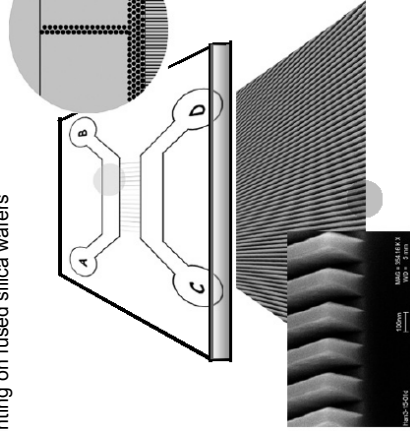
Cabodi, Turner & Craighead '02



Nanobottles

Tegenfeldt et al. '04

Nanoimprinting on fused silica wafers

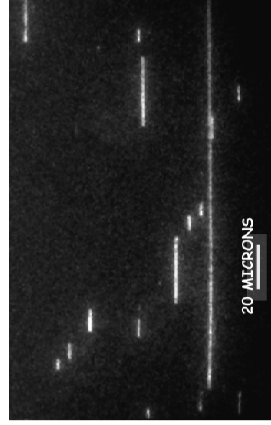


Nanobottles

Tegenfeldt et al. '04

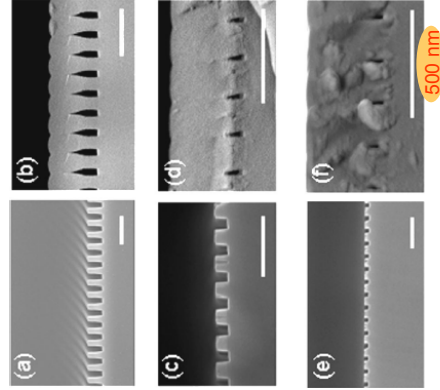
DNA stretches due to self-avoidance

$$L_z \sim L \left(\frac{bd}{D^2} \right)^{1/3}$$



Nanobottles

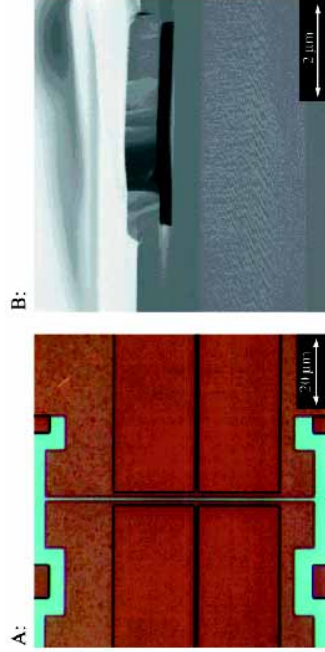
10 nm channels created by sputtering deposition



Optical sizer

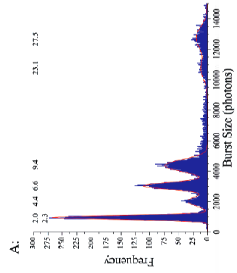
Foquet et al. '02; Chou et al. '99

Detect fluorescence from single DNA molecules flowing in a nanochannel

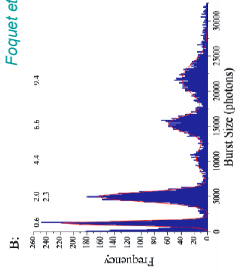


Optical sizer

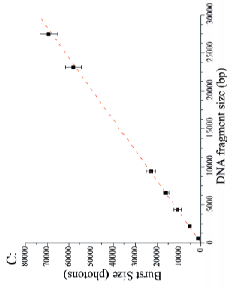
A:



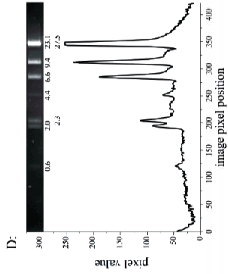
B:



C:



D:



Near-field scanner for moving molecules

Tegenfeldt et al. '00

100 nm slits illuminated from behind

