

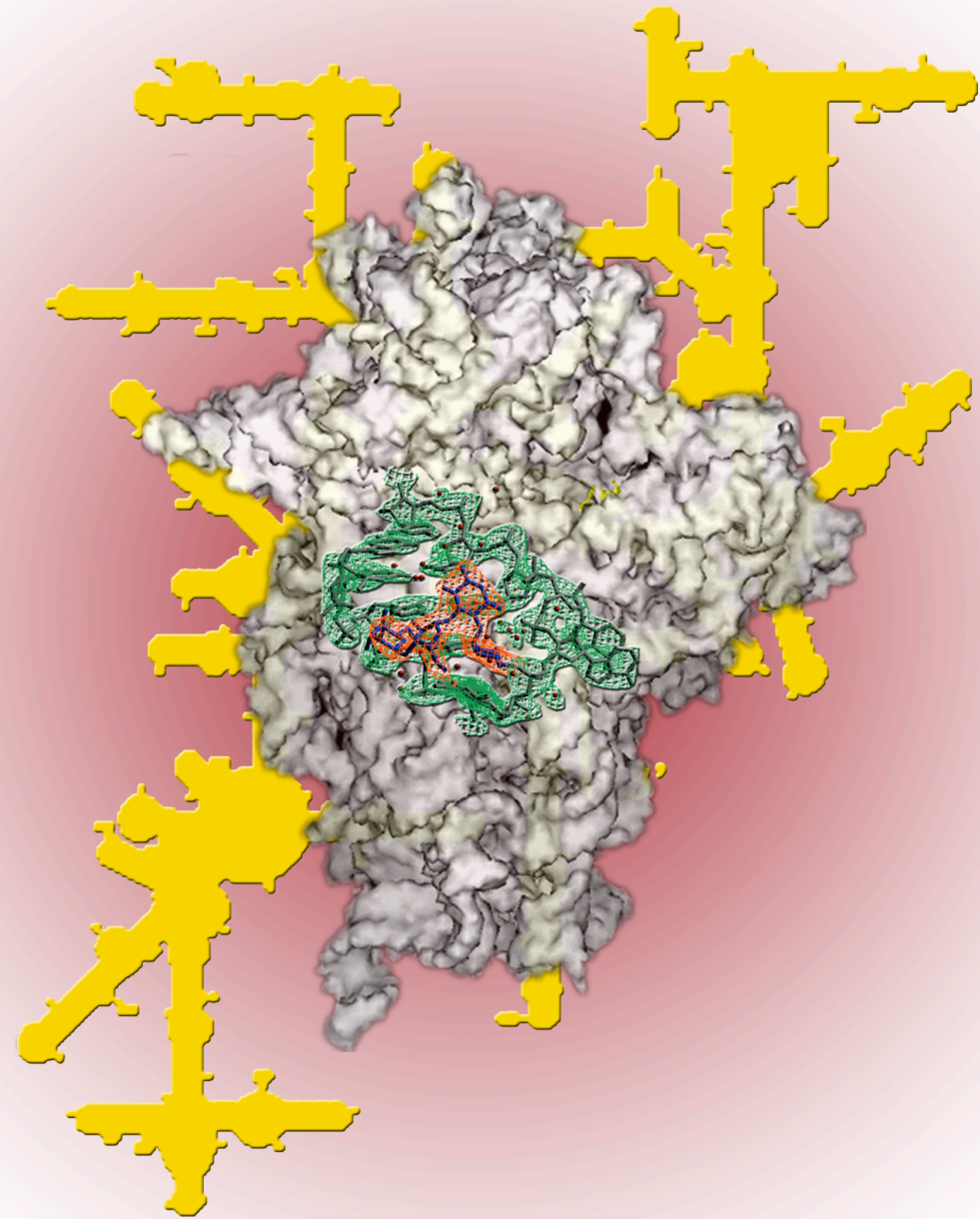
The

RNA

World IV

E. Westhof

<http://www-ibmc.u-strasbg.fr/upr9002/westhof/>

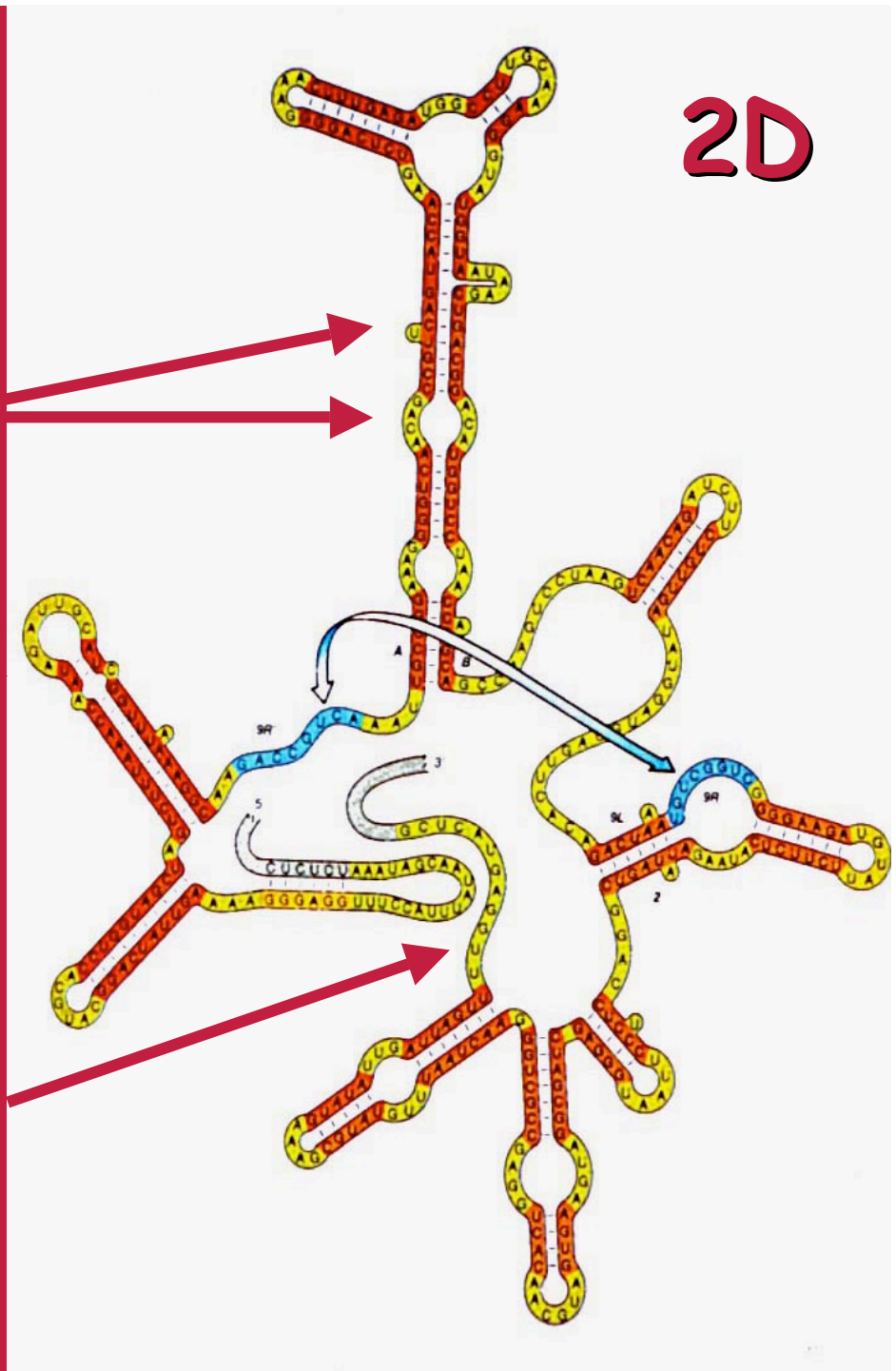


Watson-Crick base paired helices

Internal loops (symmetric, Asymmetric, bulge)

Hairpin loops

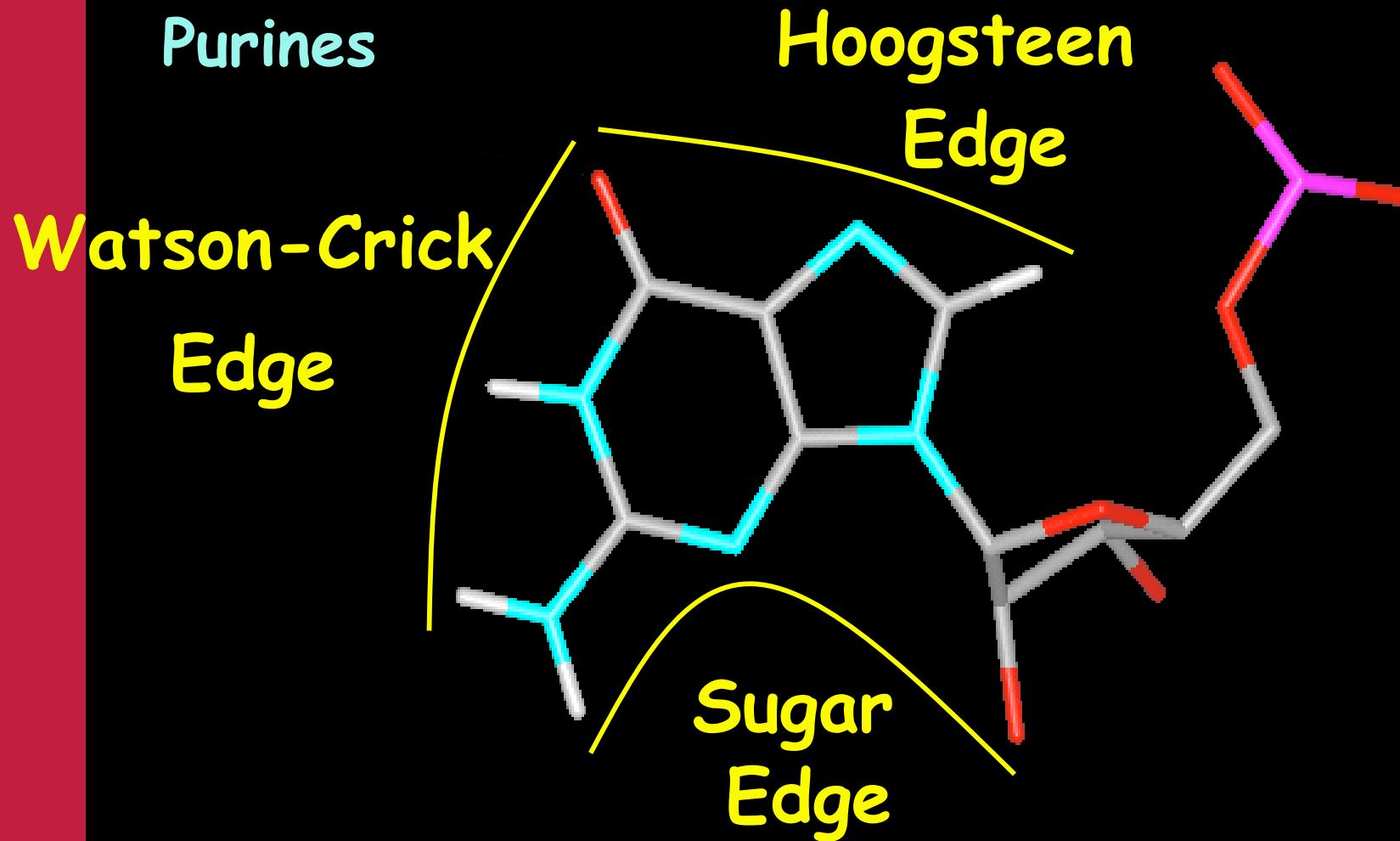
Single-strands junctions



RNA base pairing

- **Watson-Crick base pairs**
 - Form double stranded helices
 - Define the 2D structure (Main building block)
 - Dependence on monovalent ions
- **Non-Watson-Crick base pairs**
 - Form RNA motifs
 - Responsible for RNA-RNA recognition & 3D fold
 - Dependence on Divalent ions (Mg^{2+})

Three Interacting Edges



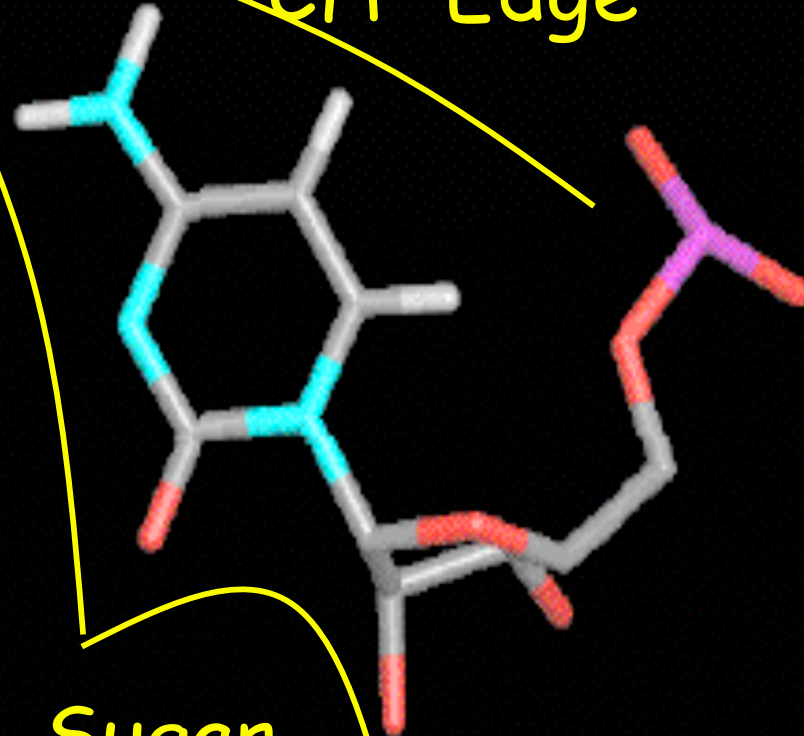
Interacting Edges

Pyrimidines

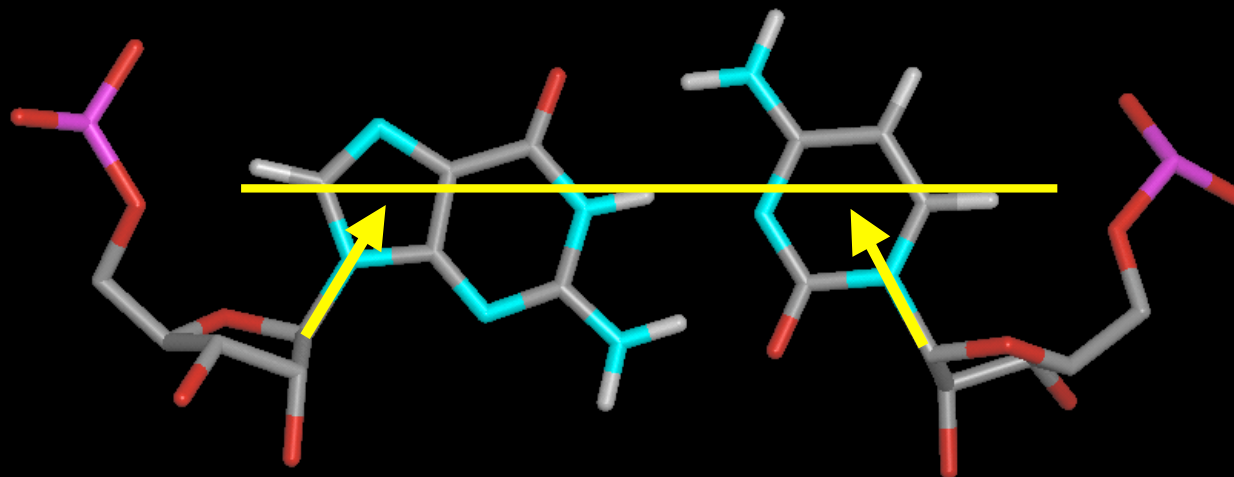
"CH" Edge

Watson-Crick
Edge

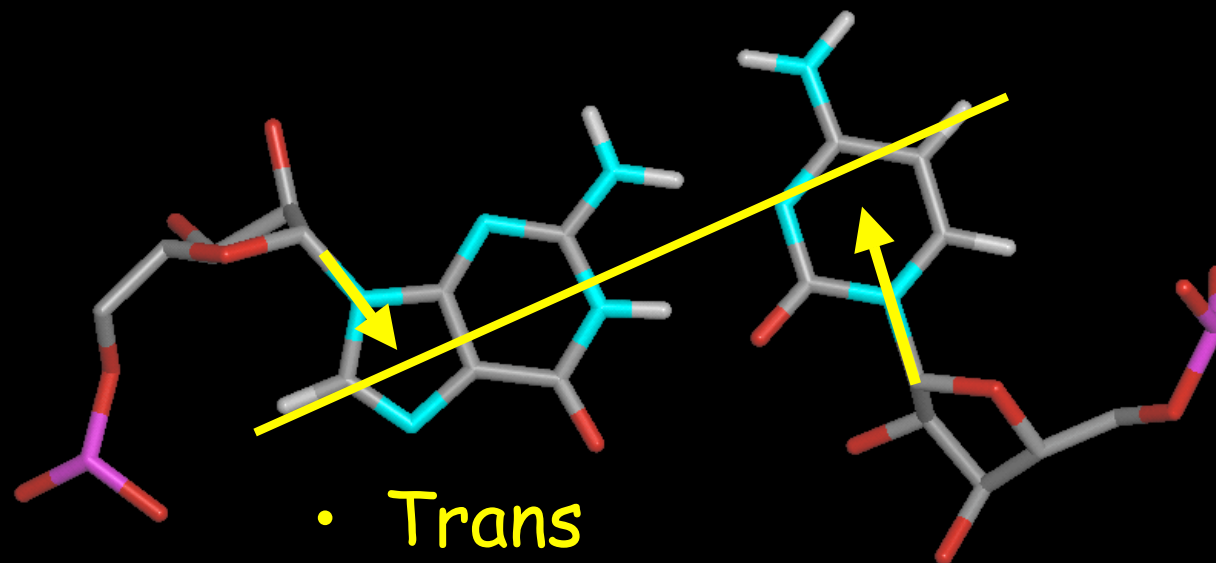
Sugar
Edge



Glycosidic Bond Orientation



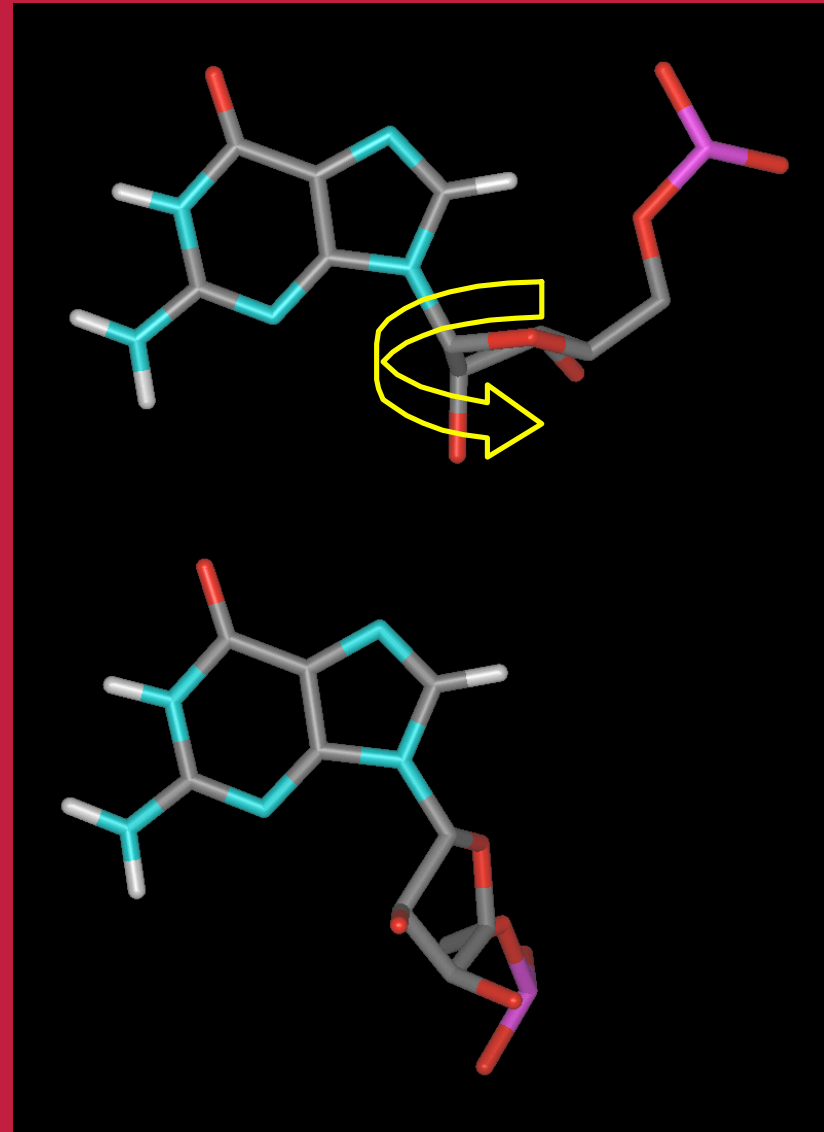
• *Cis (default)*



• *Trans*

Base-Sugar Conformation

- Anti (default)
- Syn (purines only)



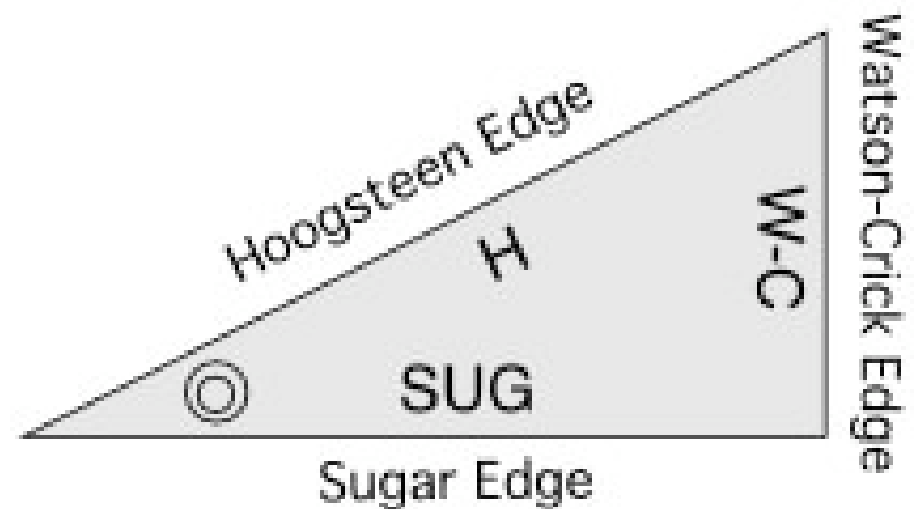
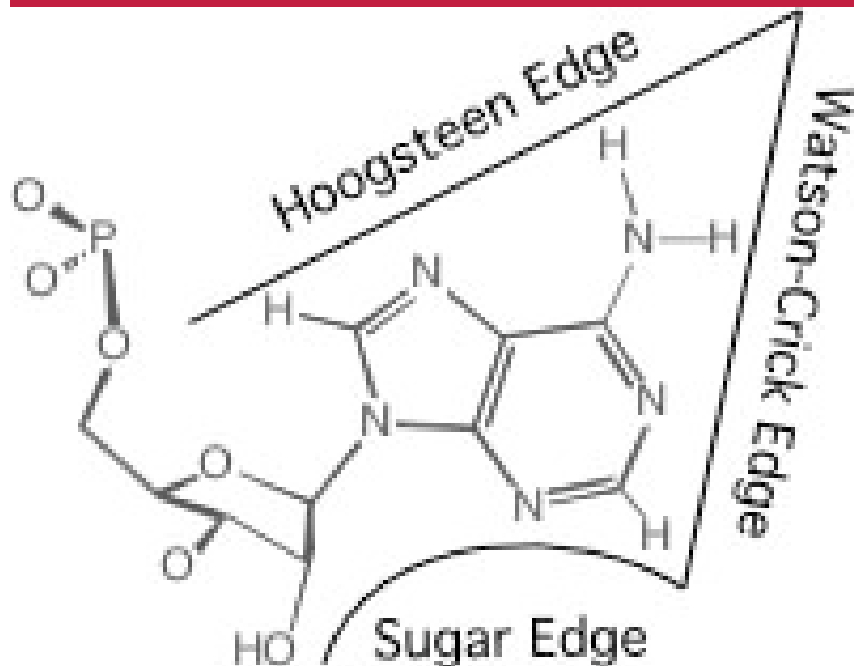
Edge-to-Edge Pairing Types

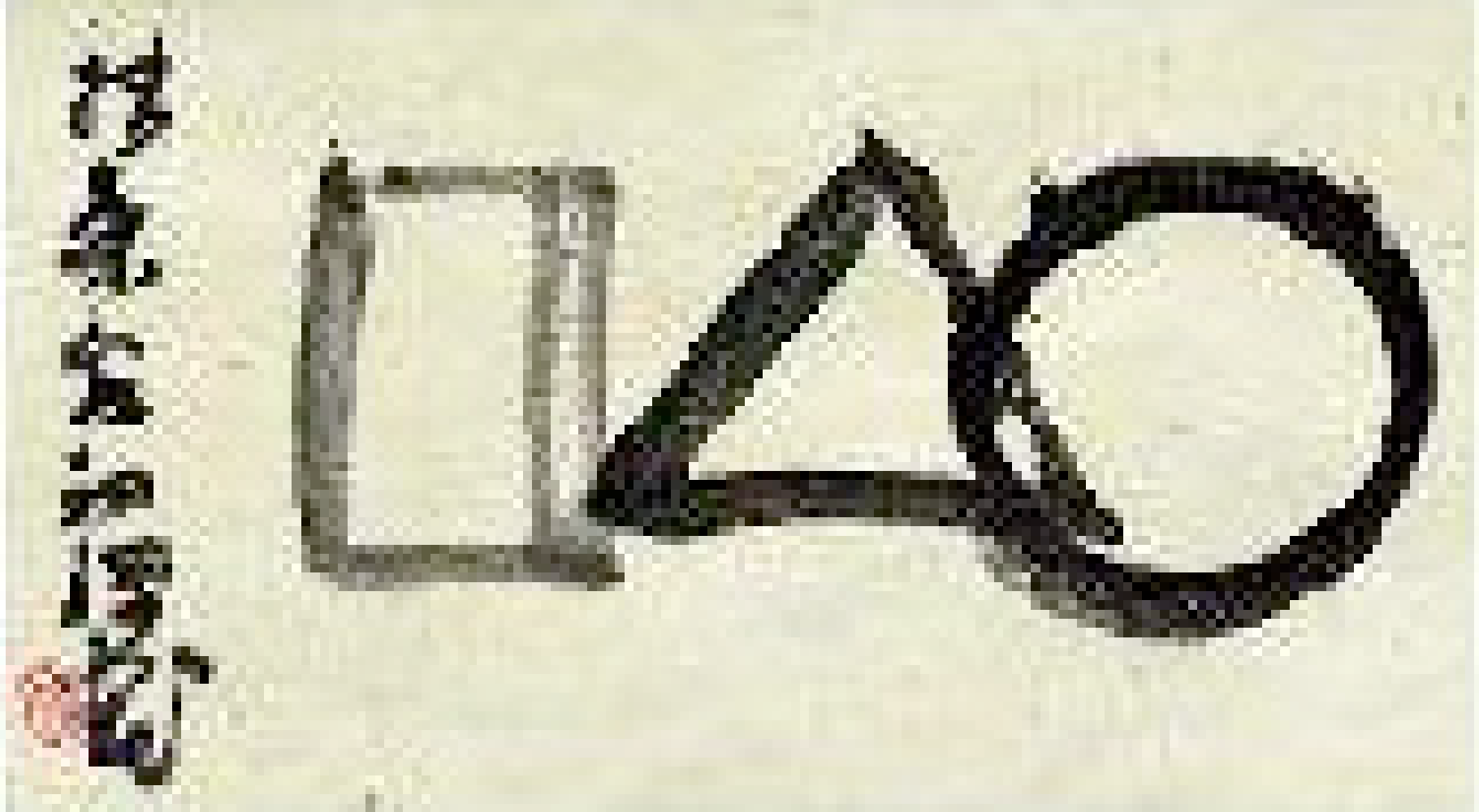


= 12 Basic Types

RNA 7, 499 (2001)

Each base has three edges
and each edge carries
various H-bonding sites

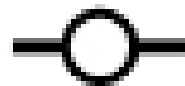




Annotations for Non-Watson-Crick Pairs



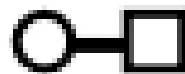
1. Cis Watson-Crick/Watson-Crick



2. Trans Watson-Crick/Watson-Crick



3. Cis Watson-Crick/Hoogsteen



4. Trans Watson-Crick/Hoogsteen



5. Cis Watson-Crick/Sugar Edge



6. Trans Watson-Crick/Sugar Edge



W.C. and Hoogsteen Pairings

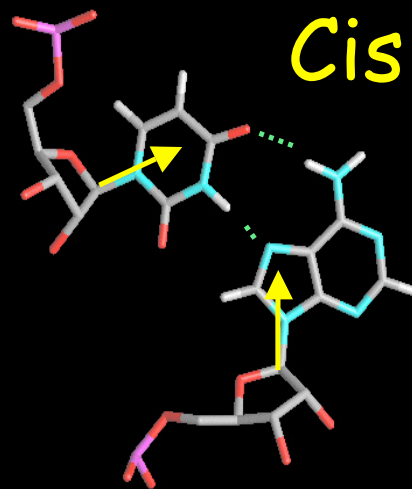
W.C.-W.C.

Cis



W.C.-Hoog.

Cis



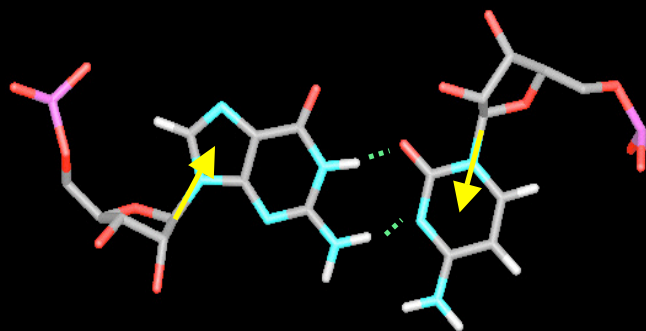
Hoog.-Hoog.

Cis

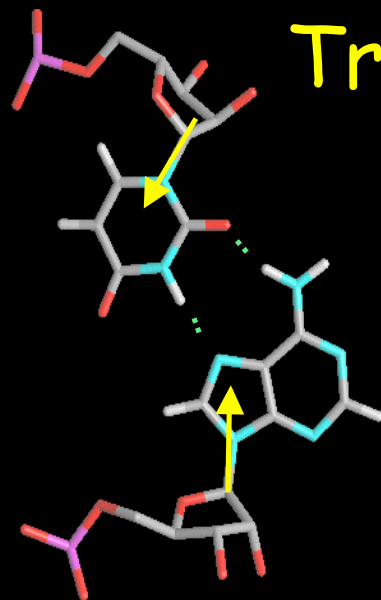
?

W.C.-W.C.

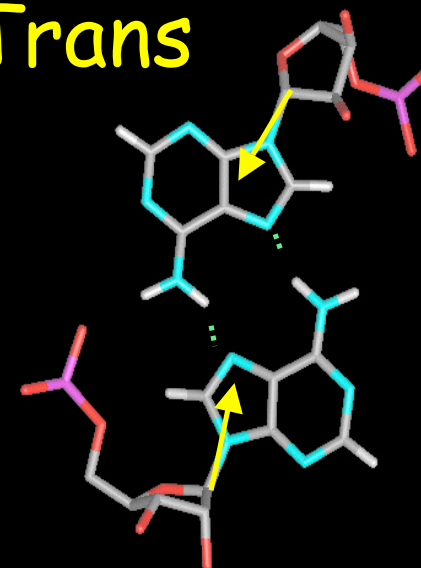
Trans



Trans



Trans

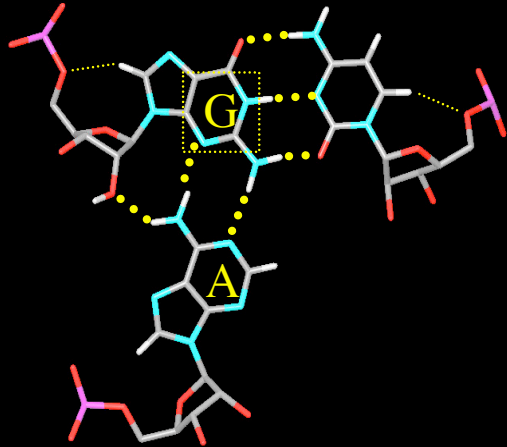


Sugar Edge Pairings

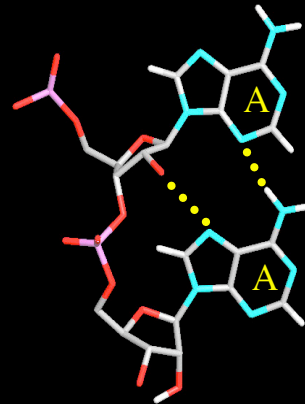
Watson-Crick

Hoogsteen

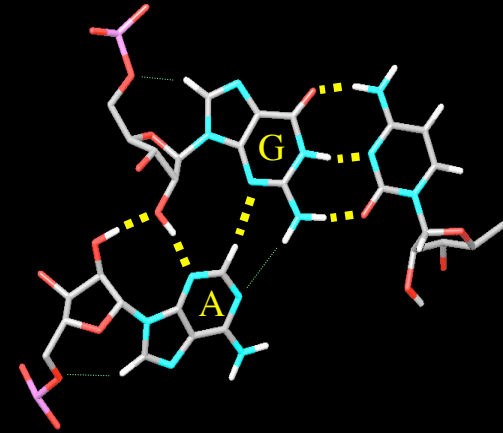
Sugar Edge



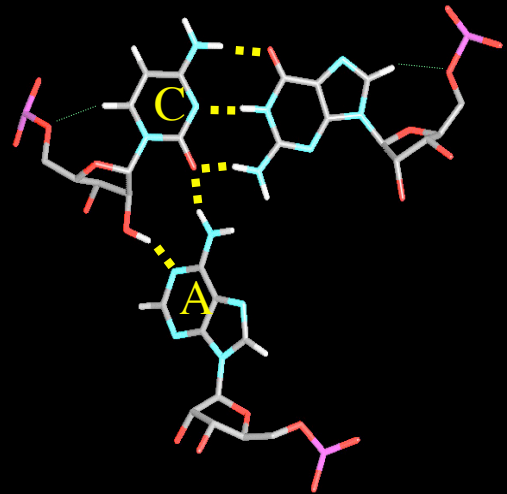
Cis



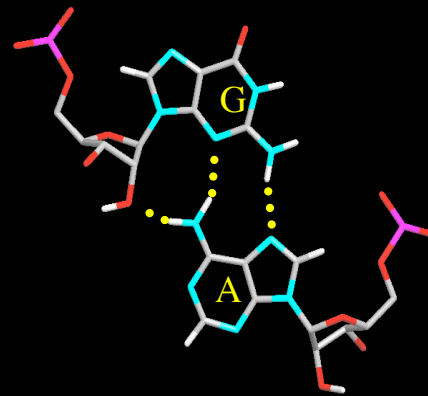
Cis



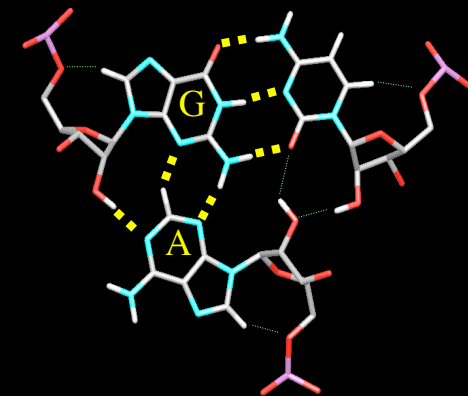
Cis



Trans



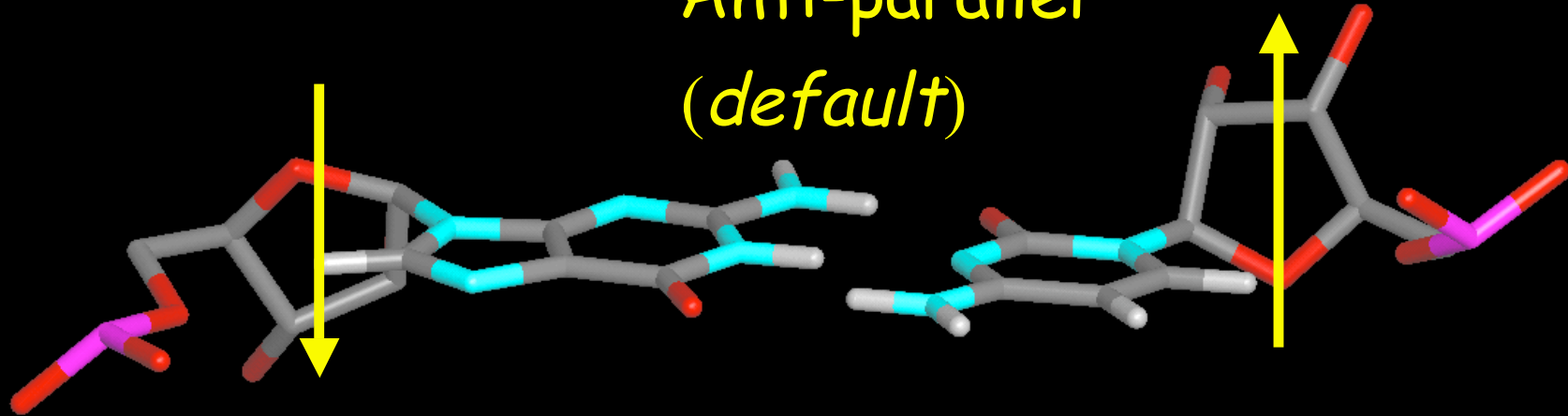
Trans



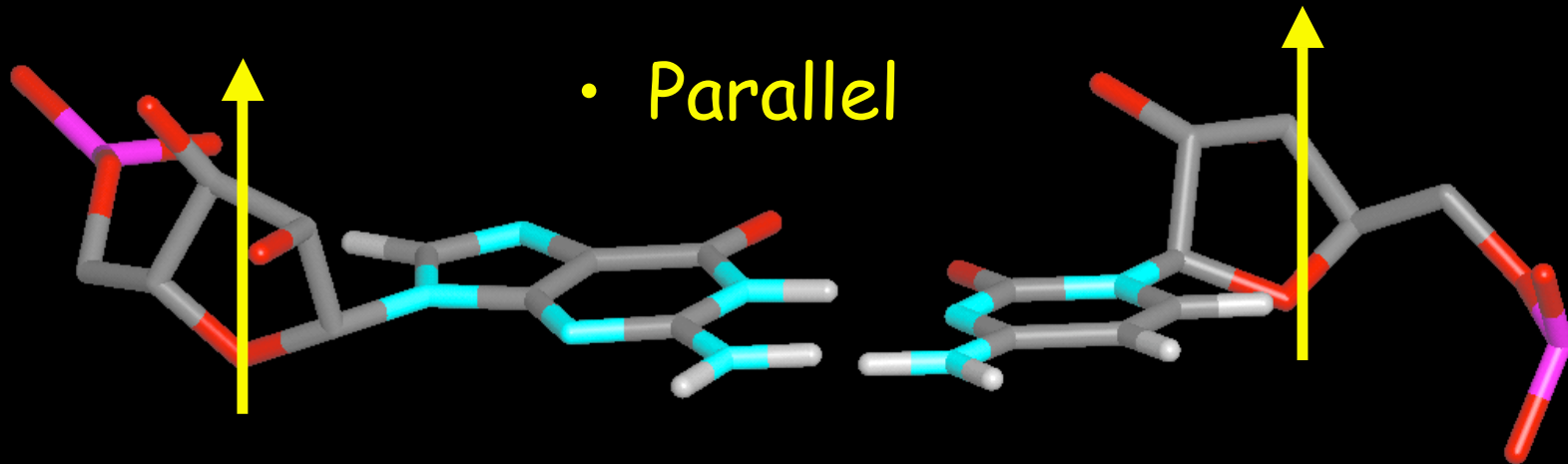
Trans

Local Strand Orientation

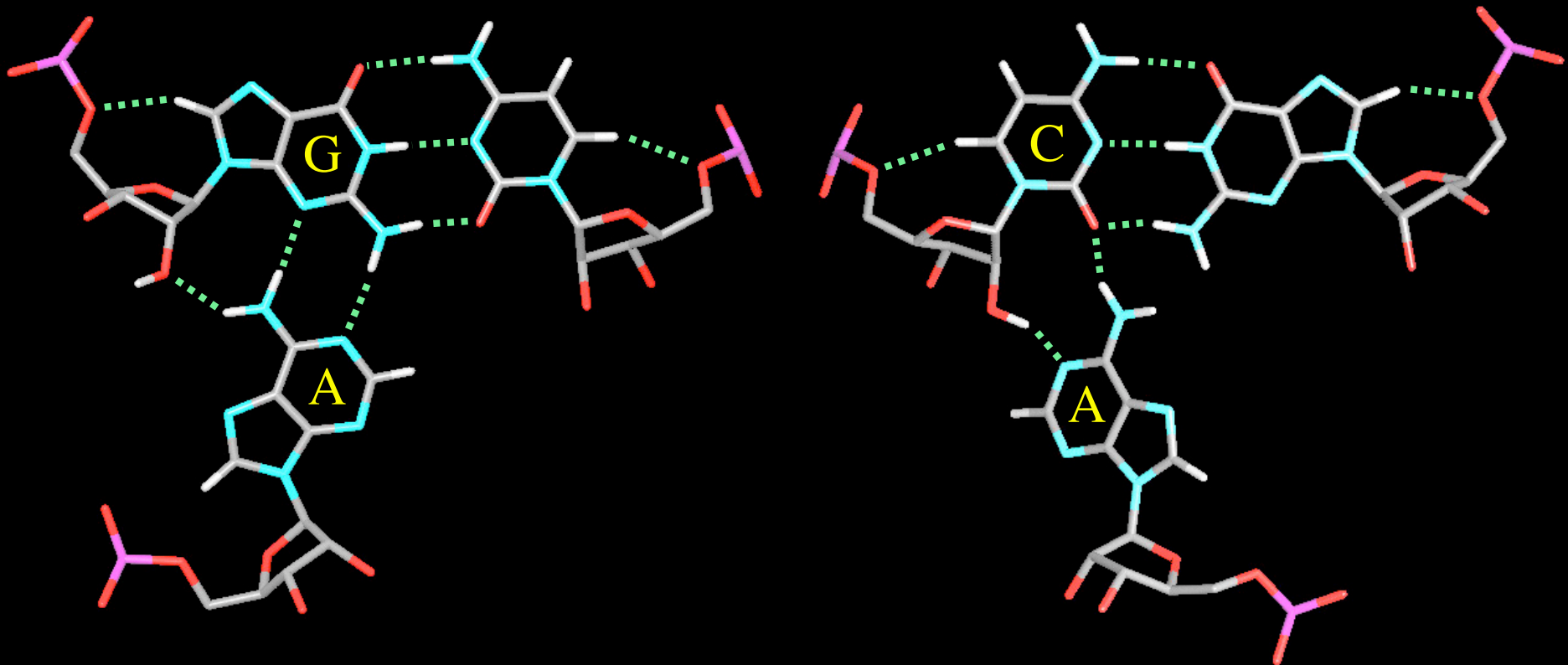
- Anti-parallel
(*default*)



- Parallel



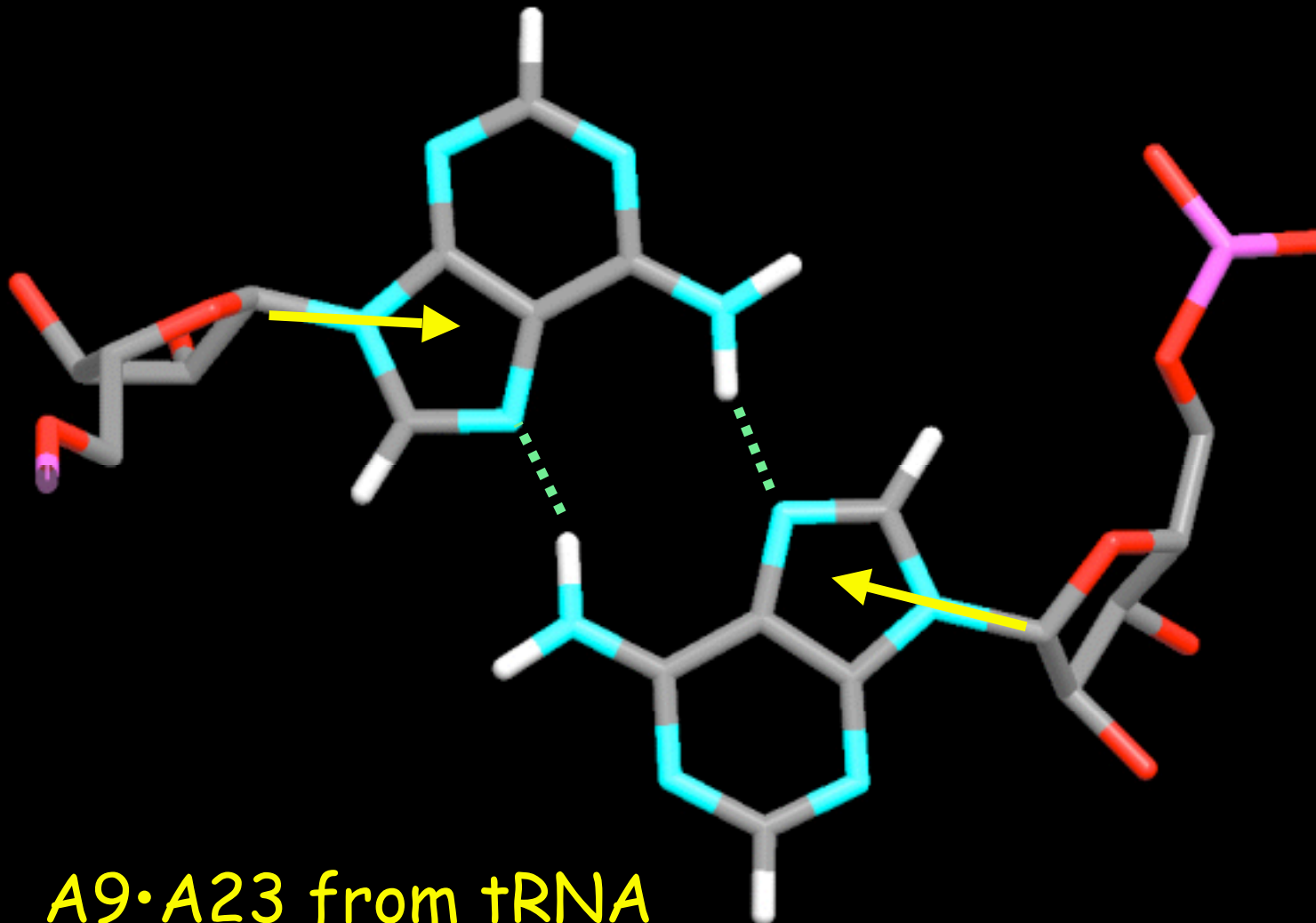
Watson-Crick/Sugar edge Pairs



Trans and Parallel

Cis and Anti-Parallel

Trans Hoogsteen/Hoogsteen



A9•A23 from tRNA

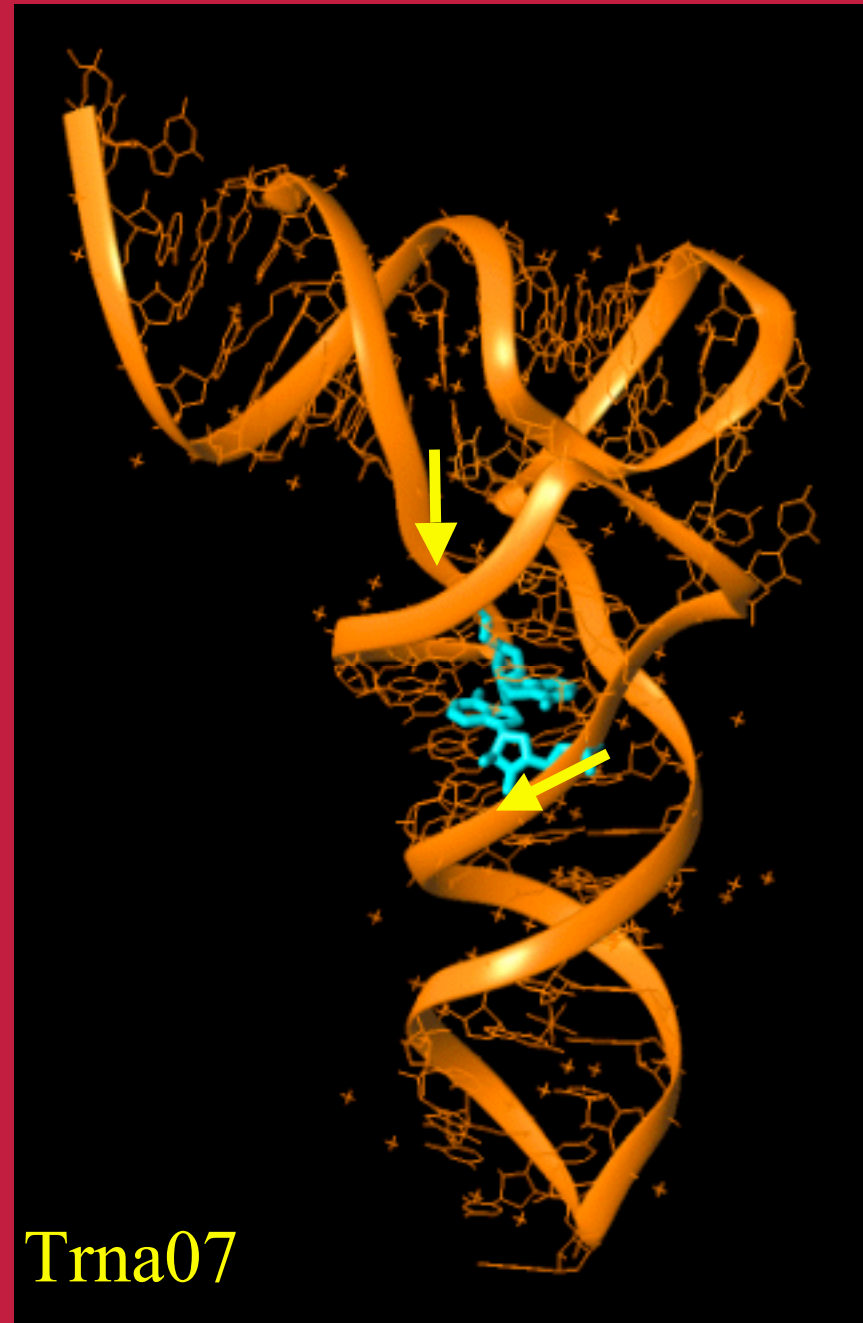
Trans Hoogsteen/Hoogsteen

Canonical A9·A23

<ul style="list-style-type: none">• Interacting Edges	<ul style="list-style-type: none">• W.C./W.C.	<ul style="list-style-type: none">• Hoog./Hoog.
<ul style="list-style-type: none">• Sugar-Base Conformations	<ul style="list-style-type: none">• Anti/Anti	<ul style="list-style-type: none">• Anti/Anti
<ul style="list-style-type: none">• Glycosidic Bond Orientations	<ul style="list-style-type: none">• Cis	<ul style="list-style-type: none">• Trans
<ul style="list-style-type: none">• Local Strand Orientations	<ul style="list-style-type: none">• Anti-Parallel	<ul style="list-style-type: none">• Parallel

Globally Parallel Strands

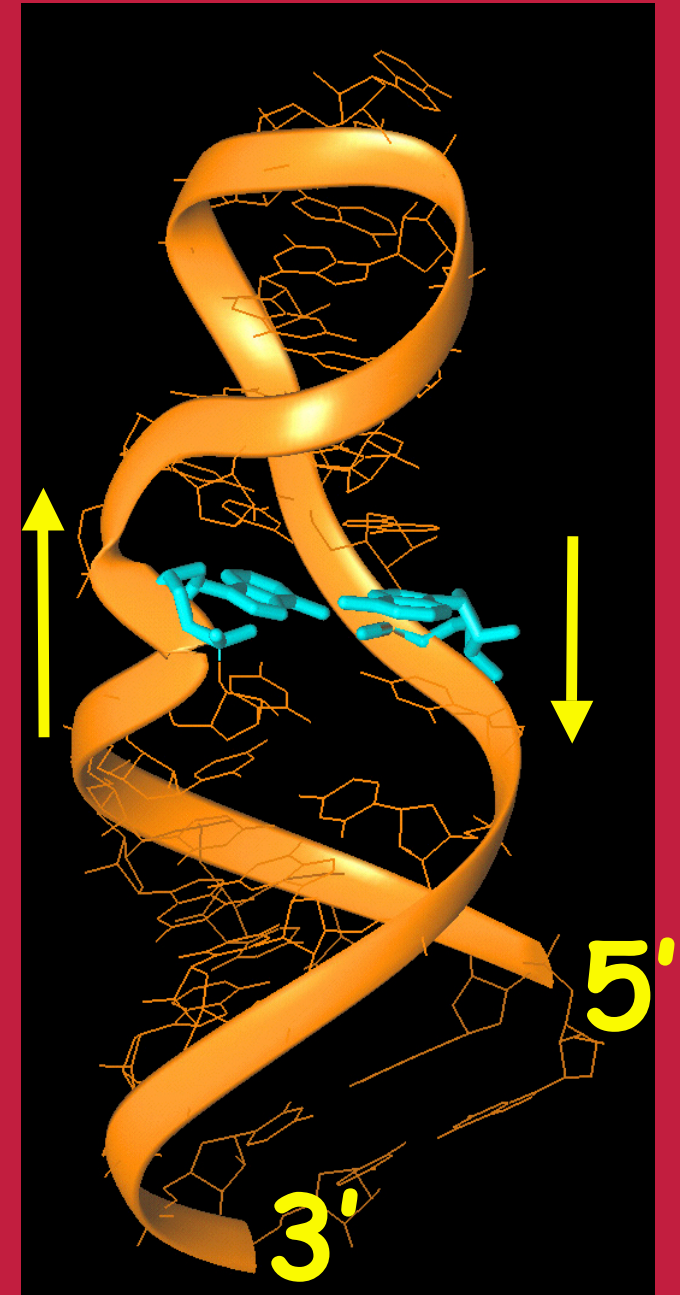
- A9•A23 in tRNA



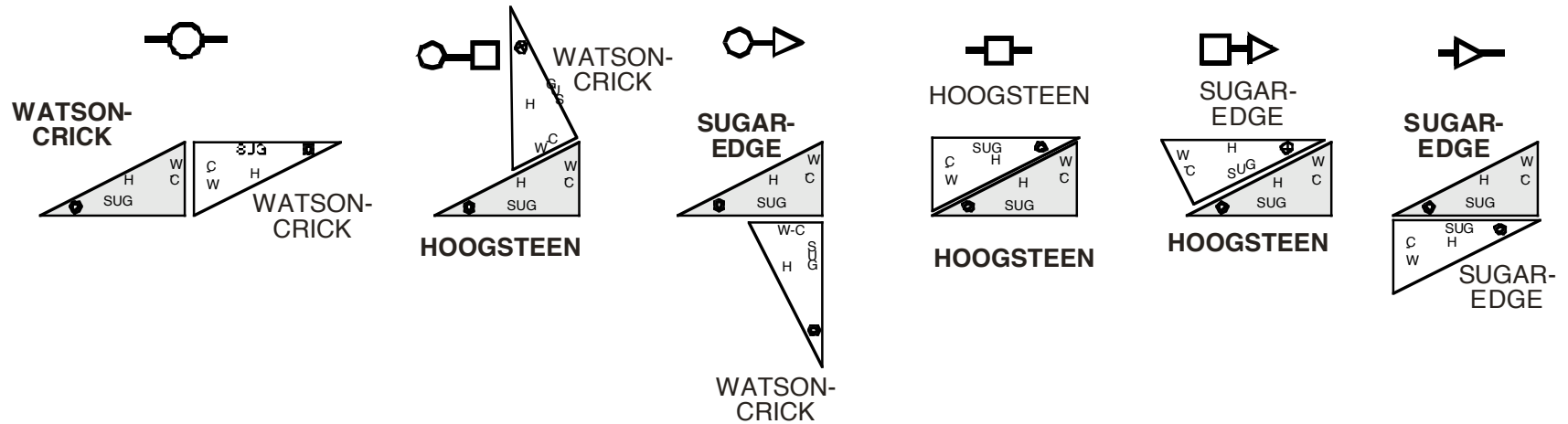
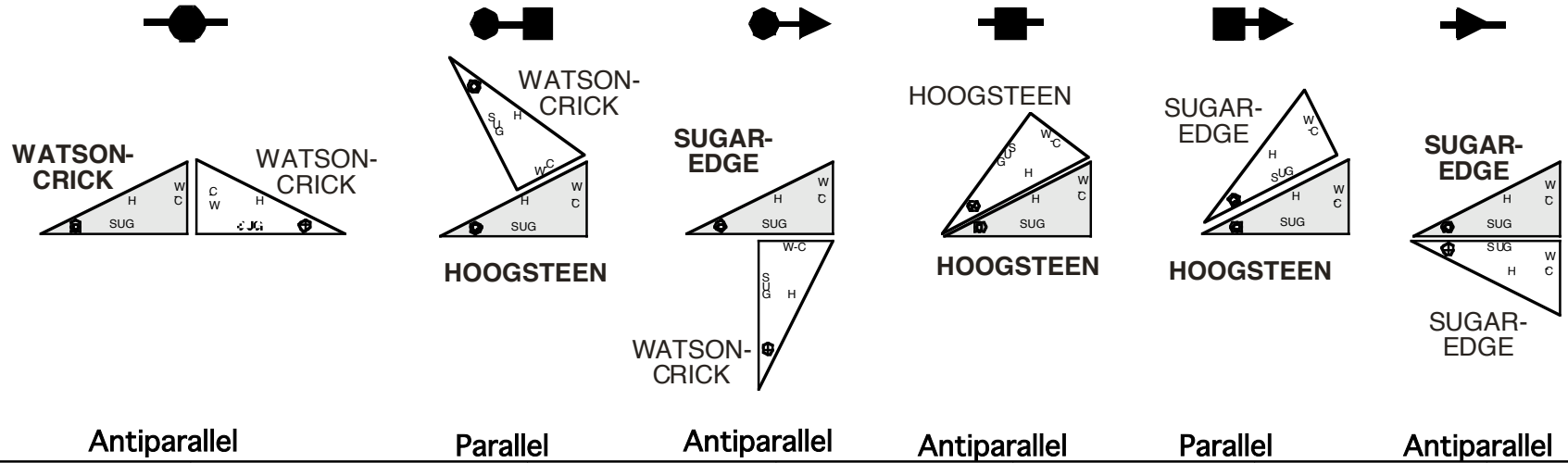
Trna07

Locally Parallel Strands

- A•A trans Hoogsteen/Hoogsteen pair in 23S rRNA "sarcin loop"
- Kink occurs next to bulged base
- Strands are globally Anti-parallel

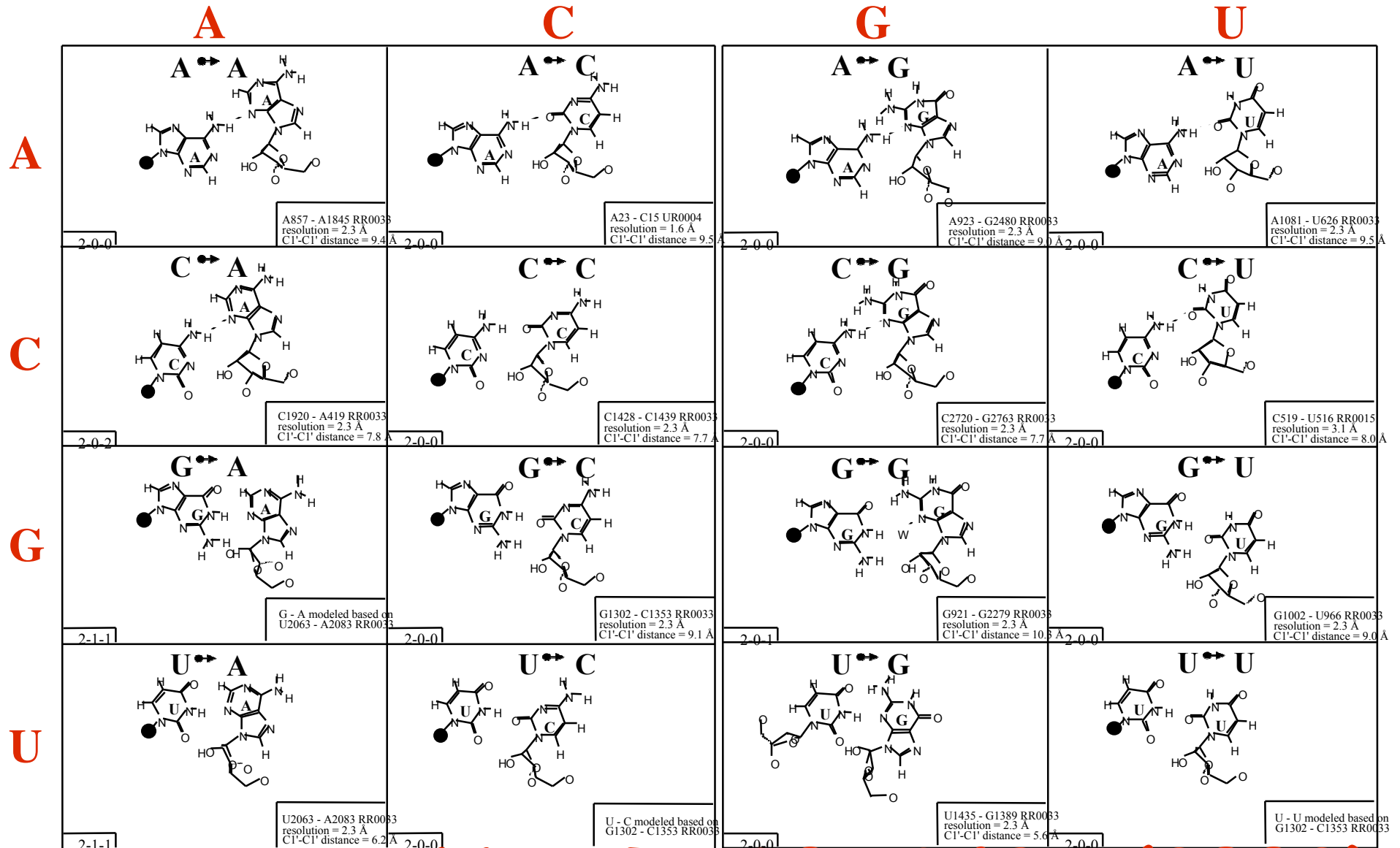


CIS BASEPAIRS



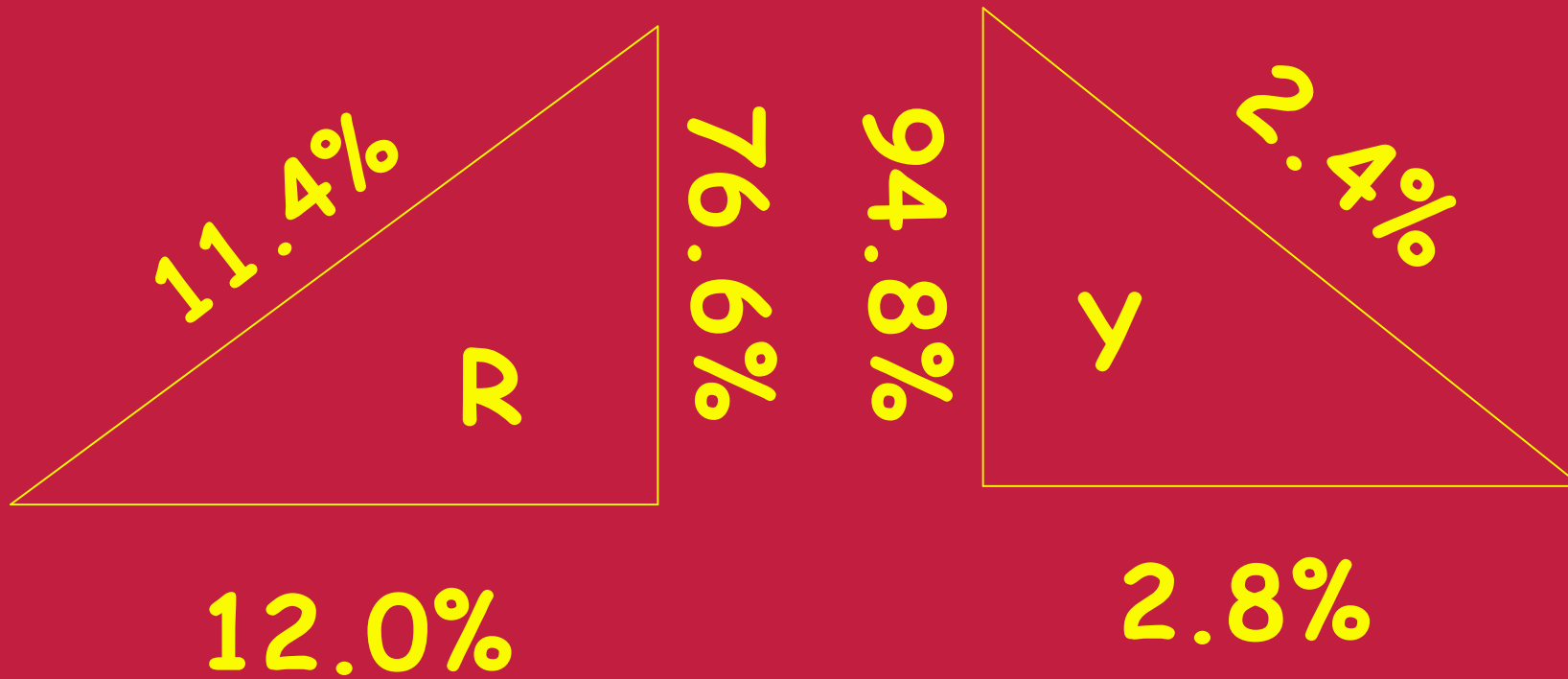
TRANS BASEPAIRS

4x4 matrix of cisWatson-Crick/Sugar pairs



N.A.R. 30, 3497 (2002)

Some statistics ...



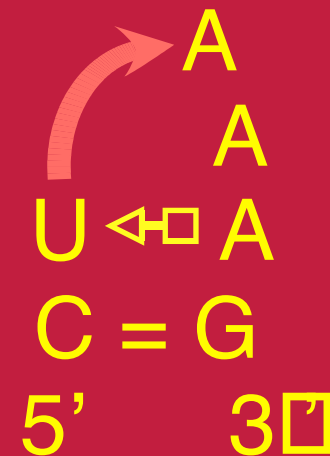
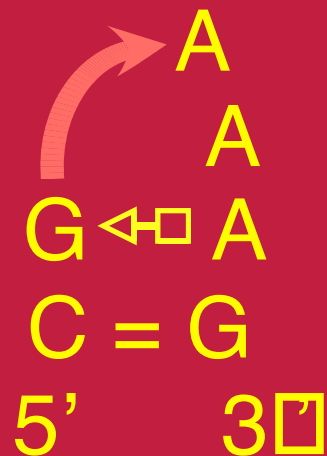
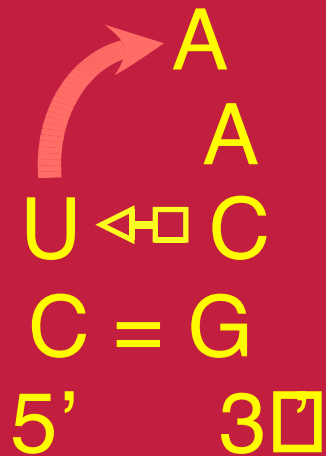
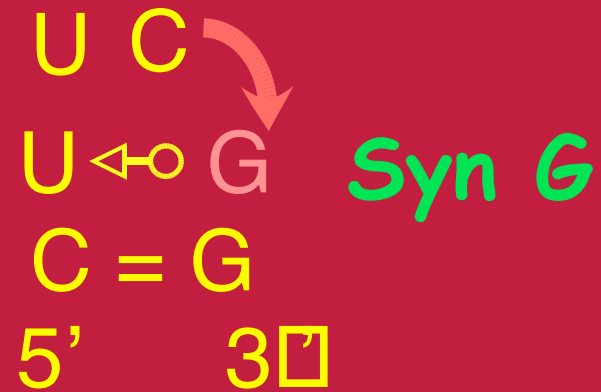
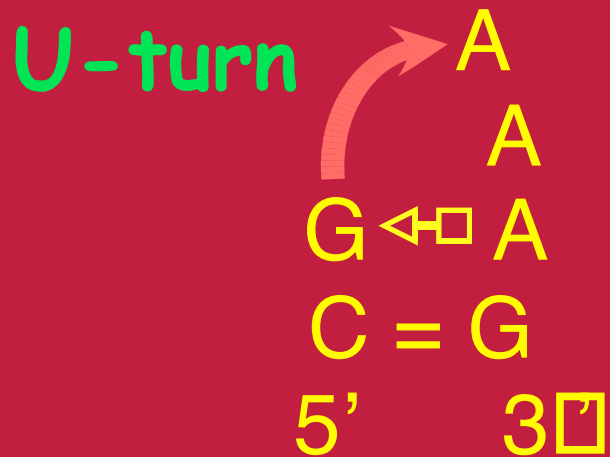
Sugar edge base pairs amongst
the most frequent non-Watson-
Crick pairs in RNA

They usually implicate the
hydroxyl O2'H

Advantages of the nomenclature

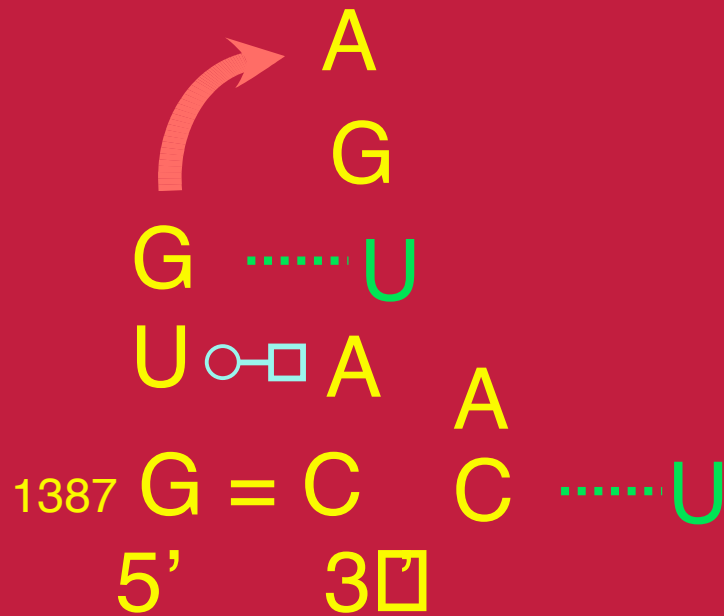
- Based on broad and defined geometrical concepts (edges) and not solely H-bonds
- Dynamical nature of molecules 'frozen' in crystals
- No crystal is perfect (data errors, resolution,...)

Diversity & Similarity in Tetraloops

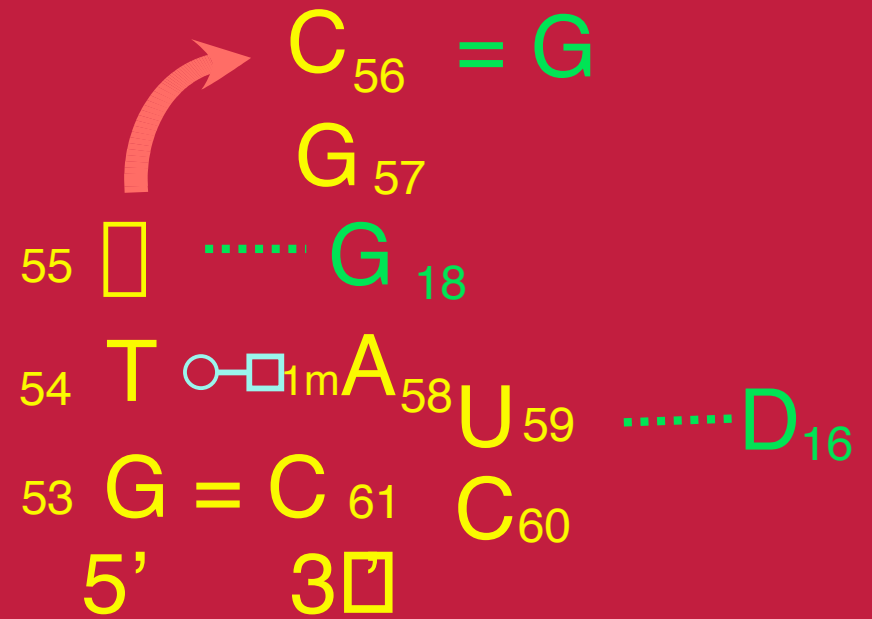


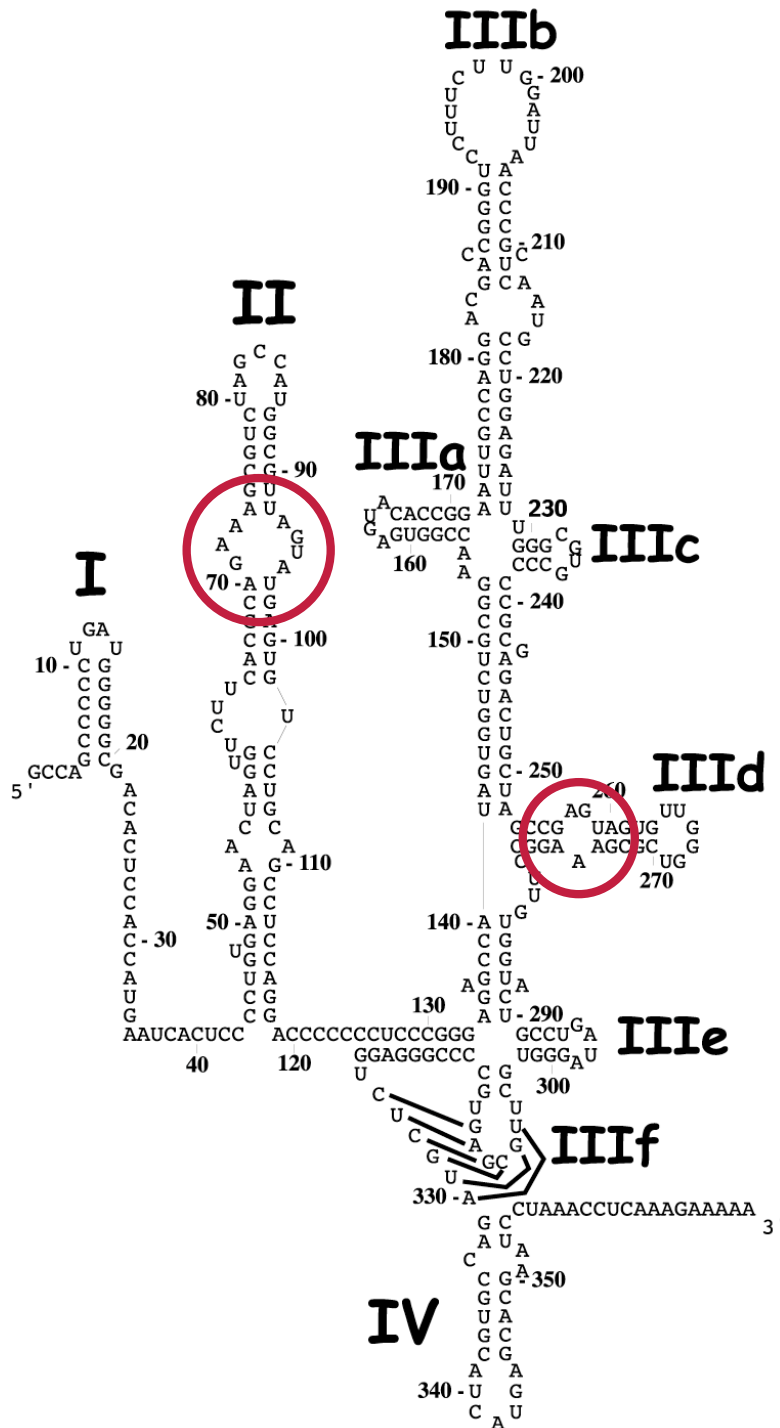
T-loops

23S rRNA



tRNA





How can we detect the presence of a motif in a given RNA ?

How can we compare motifs ?

What is a RNA motif ?

1. The U-turn ?
2. A GoA base pair ?
3. The sarcin loop ?
4. -AAUAAA- ?
5. All of those ?

**Why do we bother about
classification ?**

What are our aims ?

- 1) Derive accurate & meaningful alignments;
- 2) Extract from a set of 'aligned' sequences, information about 3D contacts and architecture;
- 3) Derive rules for RNA evolution;
- 4) Search genomes for non-coding RNAs.

A MOTIF is an ensemble of ordered elements under constraints.

Sequential motifs :

- Strict : -AUG-
- Fuzzy : -AAUAAA-

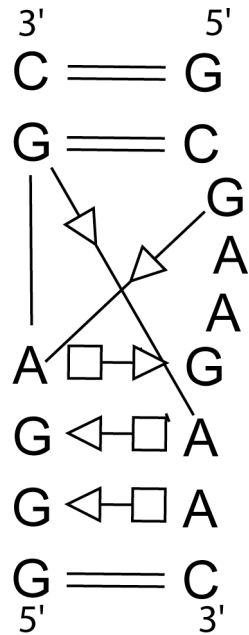
Structural motifs:

- -GNRA-
- Boxes C/D or H/ACA

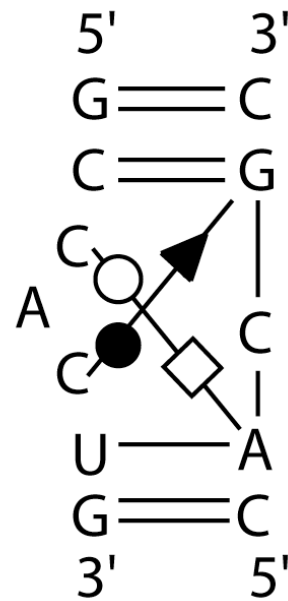
Consequences of this definition :

- Must fit within helices
- Should be recurrent
- Should they fold autonomously ?

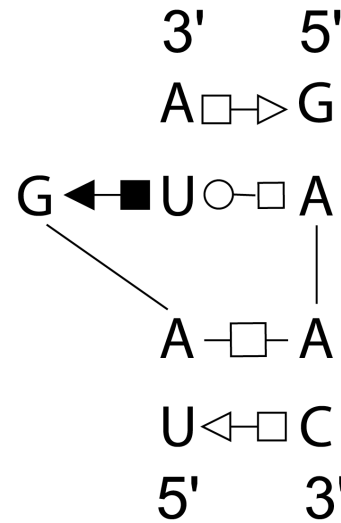
The K-turn



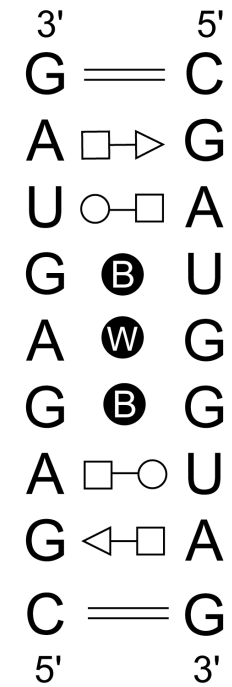
The C-motif

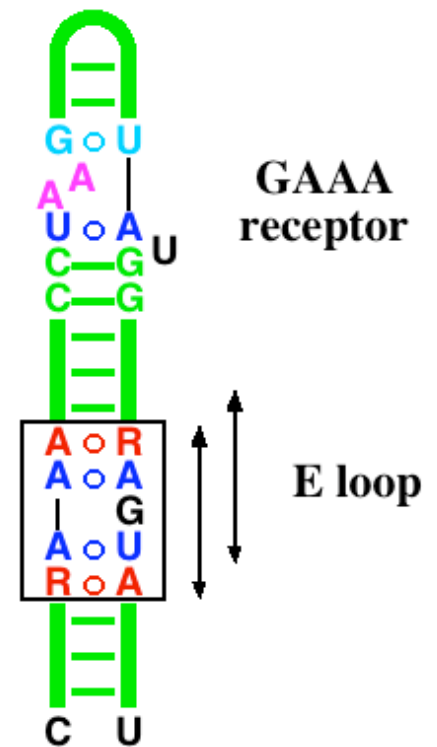
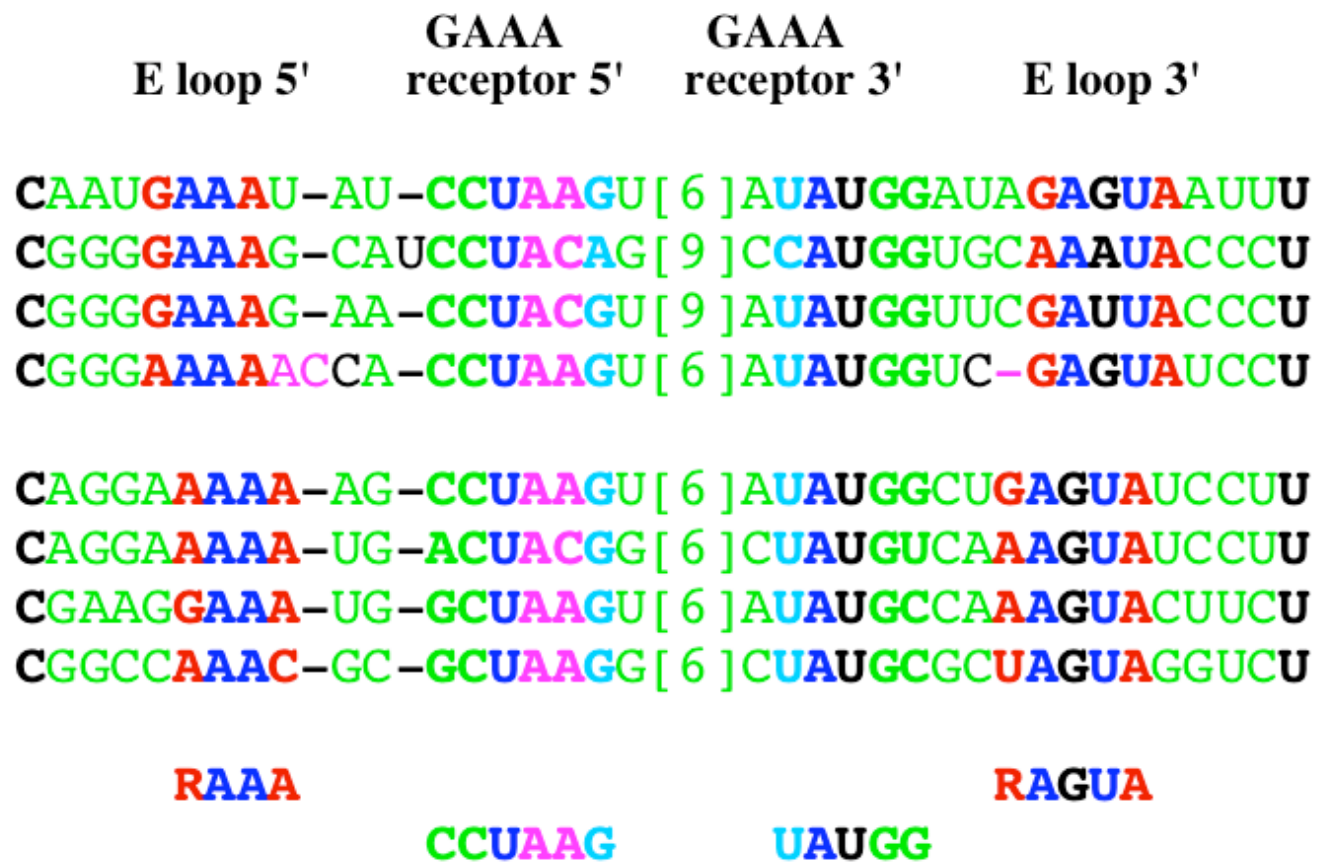


The sarcin motif



The loop-E motif





Would such motifs evolve ?

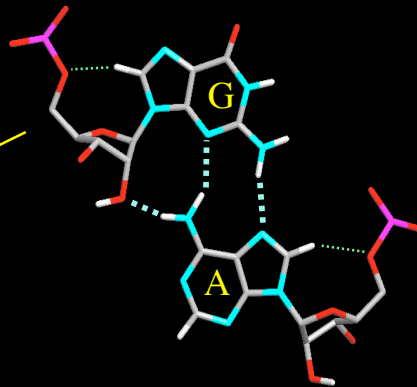
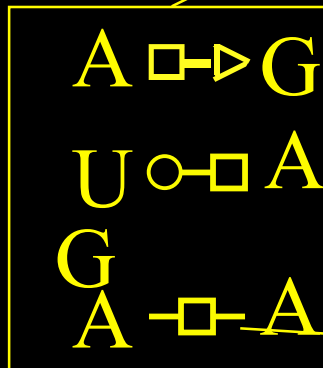
- Chemistry and geometry of base pairs constrain the evolution of motifs (> structural convergence)
- Such motifs are spread across the Tree of Life and are used for diverse functions
- Such motifs cannot be used to infer homology

S-Motif Base-pairing

Trans Hoog./Sug.

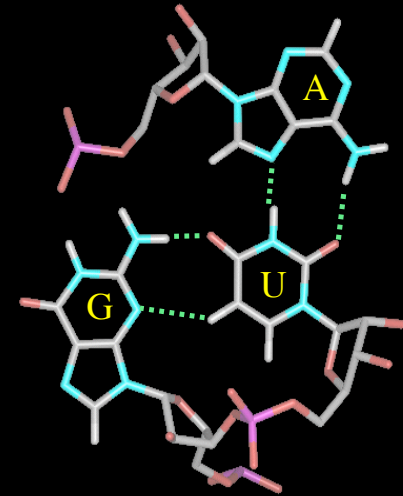
A·G

3' | 5'



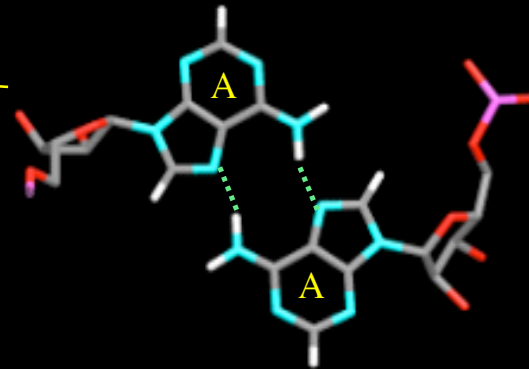
Trans W.C./Hoog.

U·A



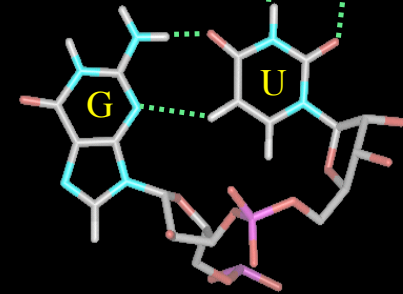
5' | 3'

Trans Hoog./Hoog. A·A

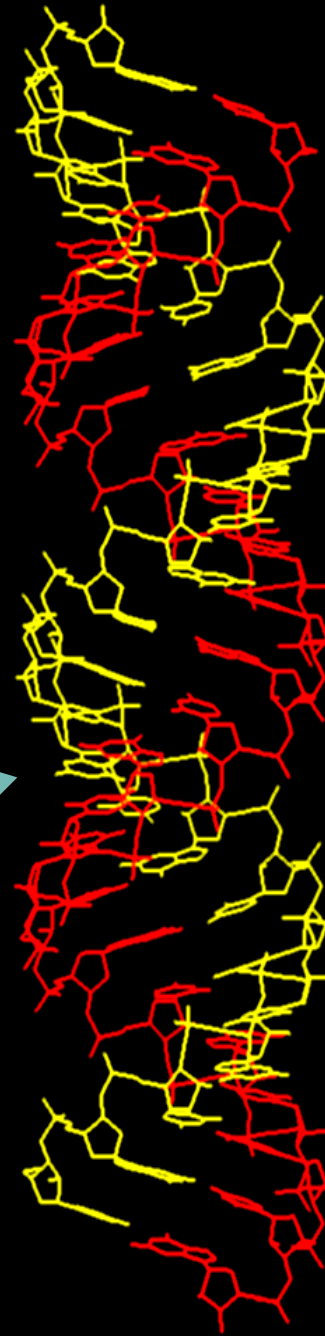
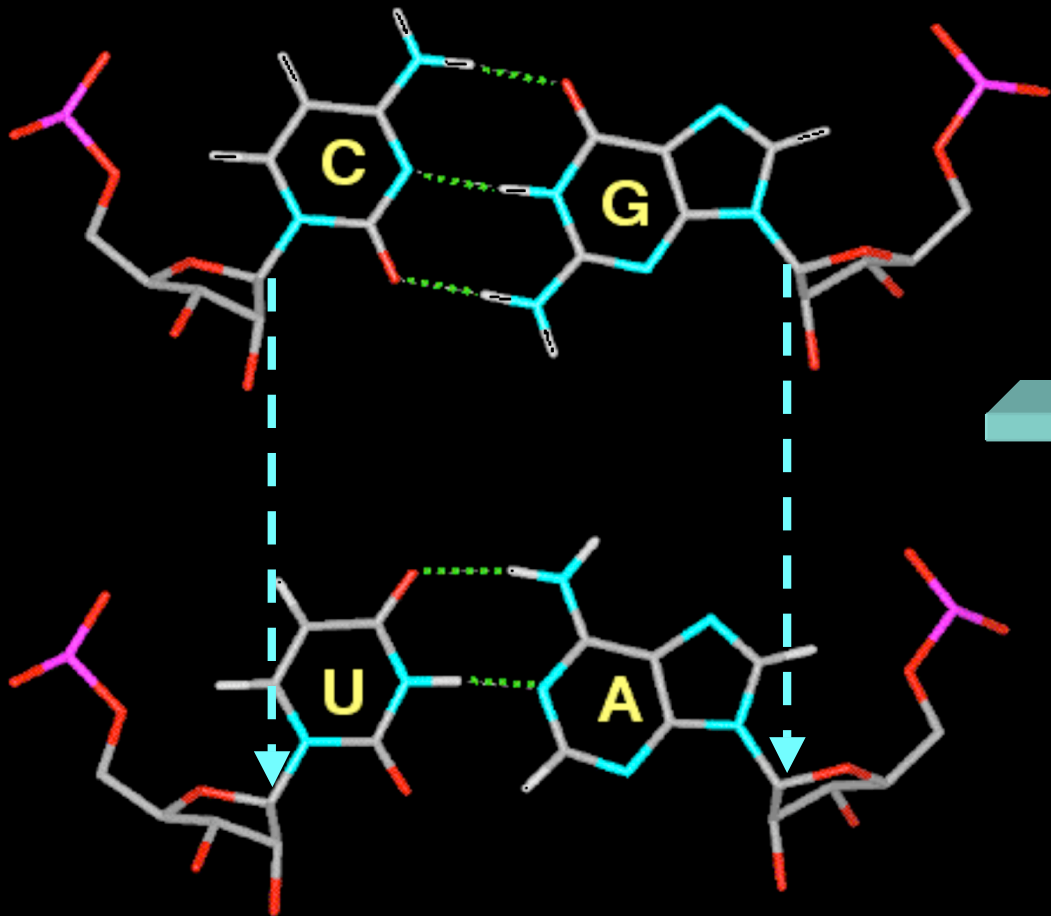


Cis Hoog./Sug.

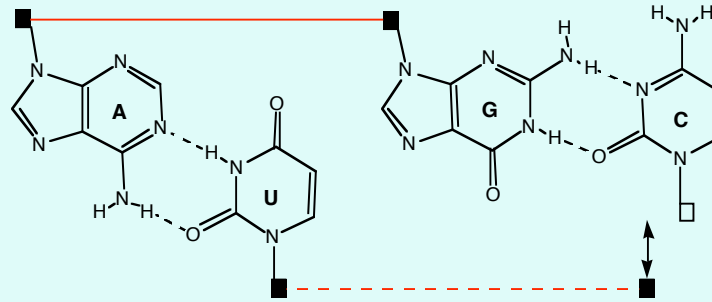
U·G



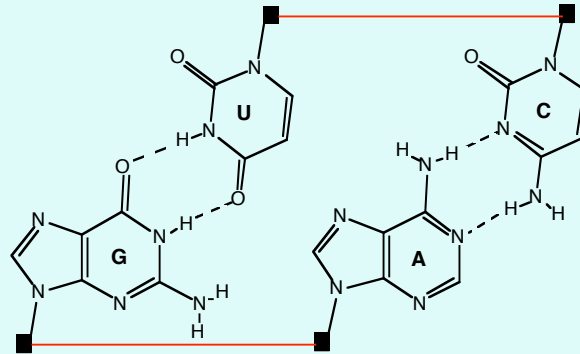
Watson-Crick pairs are isosteric



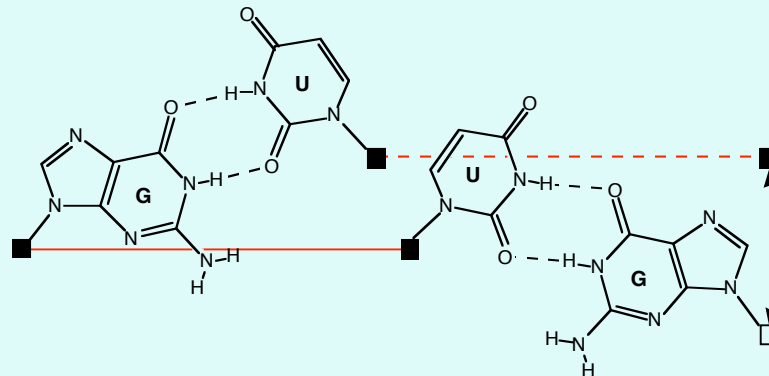
Non-isostérie des appariements Watson-Crick trans pour les bases complémentaires.

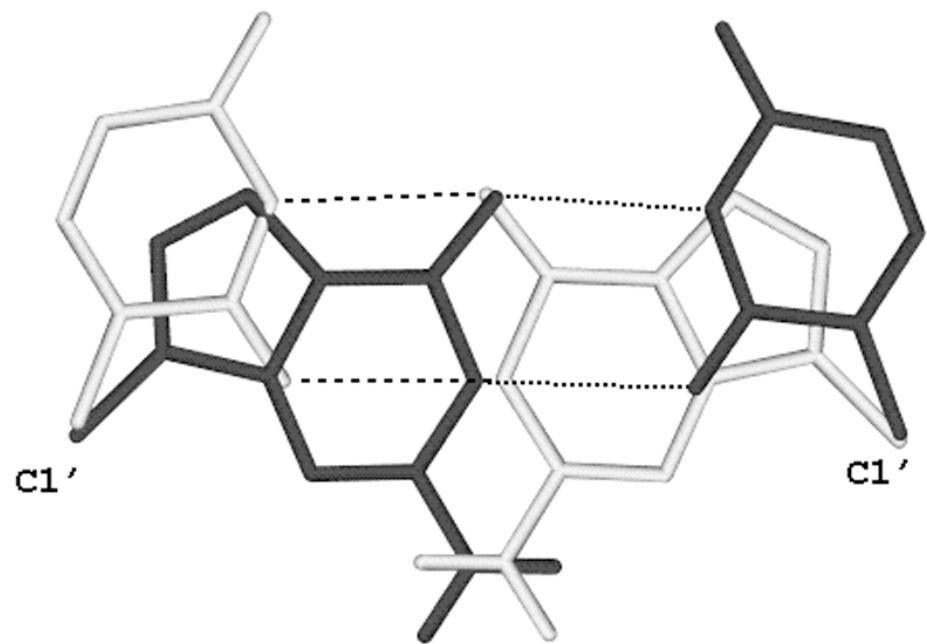
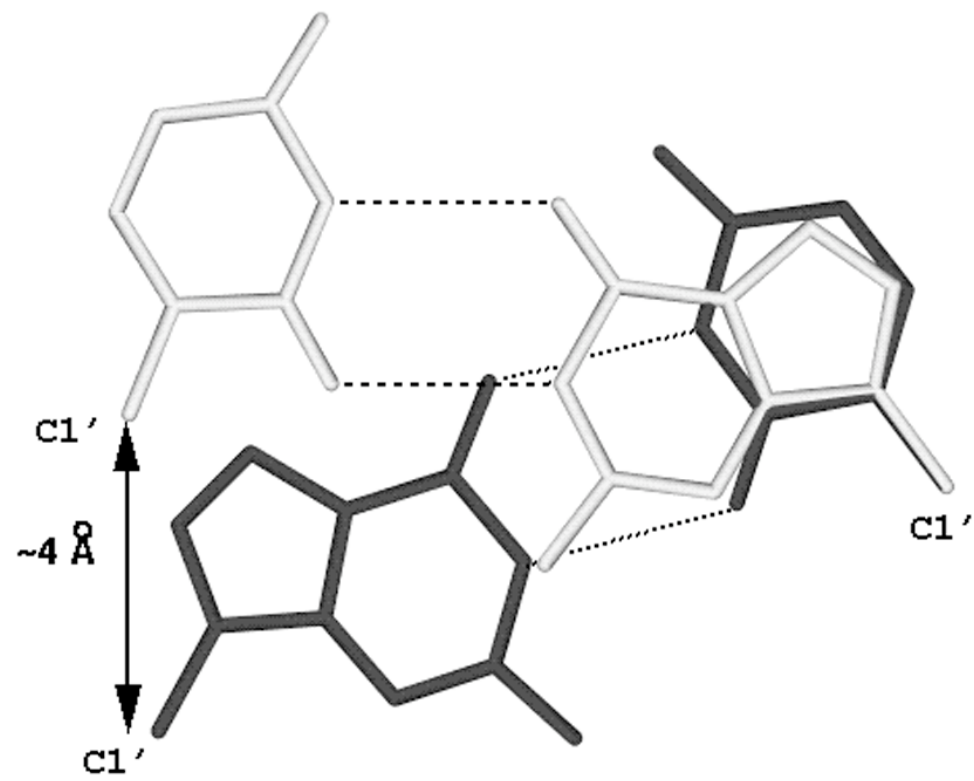


Isostérie des appariements Watson-Crick trans G-U et A-C

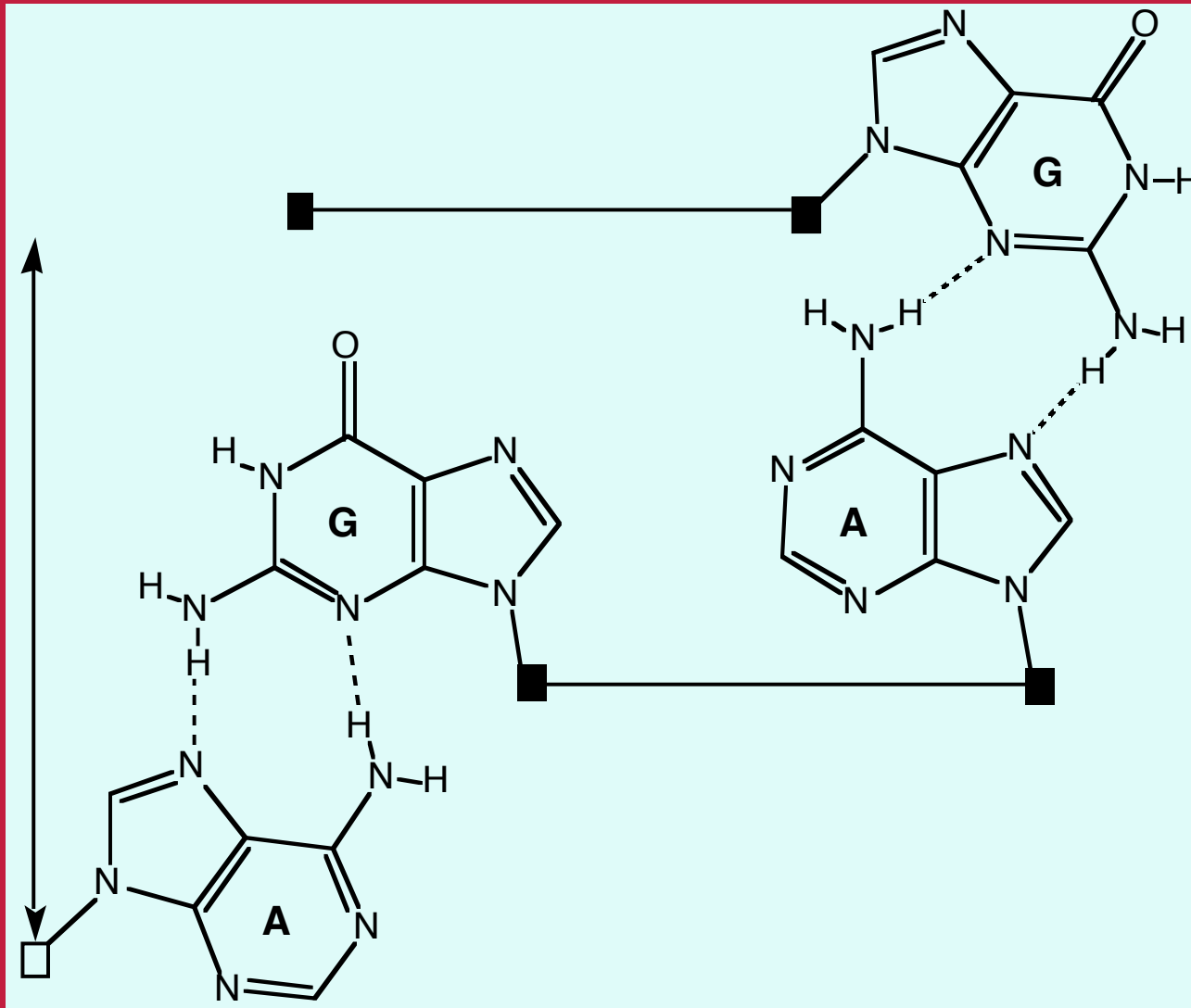


Non-isostérie des appariements Watson-Crick cis G-U et U-G





Non-isostericity of Hoogsteen/Sugar base pairs (or sheared AoG pairs)



Subsets of isosteric pairs

- Each of the 12 geometric families comprises 4x4 combinations (16x12=192)
- Only some are chemically allowed
- Those are organized in subsets of isosteric pairs
- The isosteric pairs are revealed by correlating 3D structures with sequence comparisons and modelling

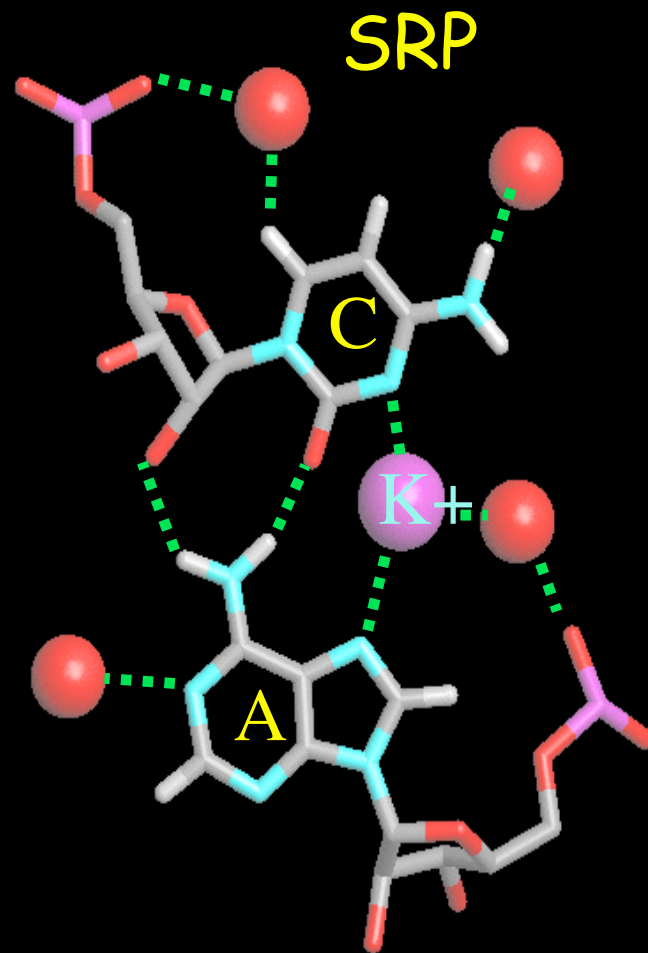
Isostericity Matrix for cis W.C./W.C.

Watson-Crick

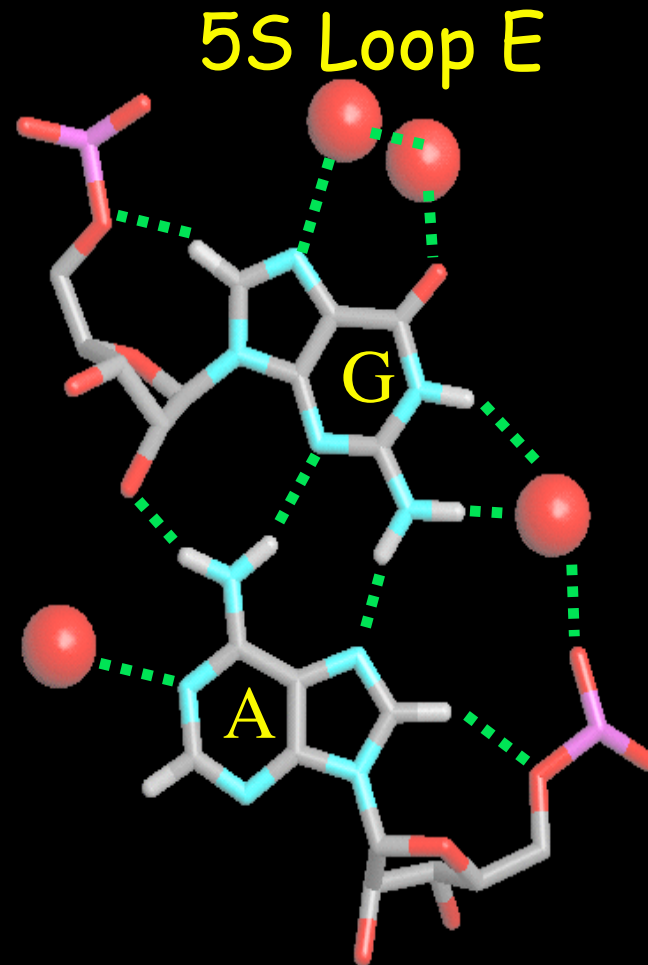
Watson-Crick

	A	C	G	U
A	I5	i3	I2	I1
C	I3	I5	I1	I4
G	I2	I1		i3
U	I1	I4	I3	I5

Trans Hoogsteen/Sugar

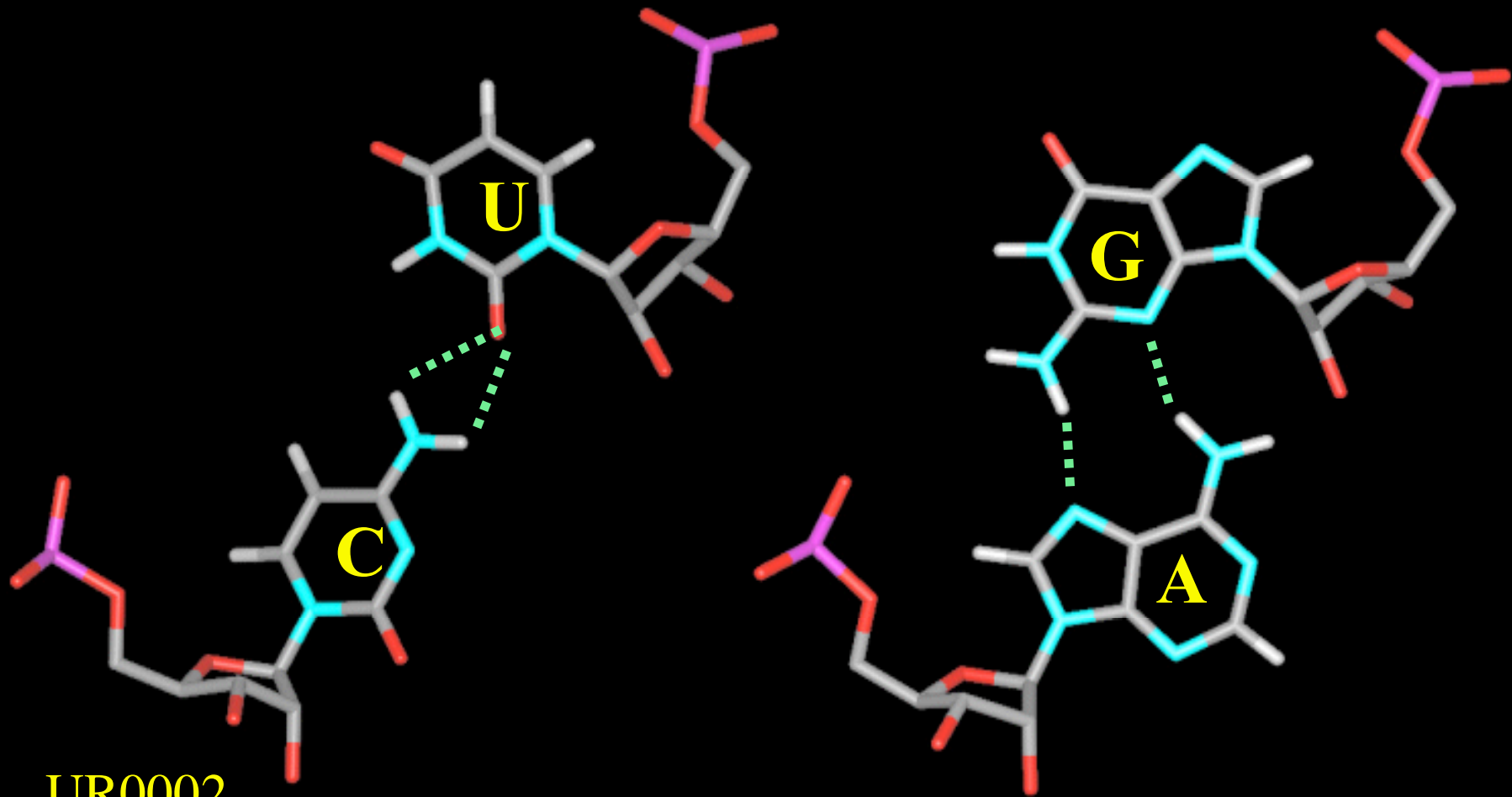


NDB file PR0021
Batey et al. (2000)



NDB file URL064
Correll et al. (1997)

C·U Isosteric to "Sheared" A·G (*trans* Hoogsteen/Sugar edge)



UR0002

Isostericity Matrix trans Hoogsteen/Sugar Edge

Sugar Edge

H
o
o
g
s
t
e
e
n

	A	C	G	U
A	I1	I1	I1	I1
C	I1	I1		I1
G			I2	
U	I2		I2	

Sequence signature of RNA folds

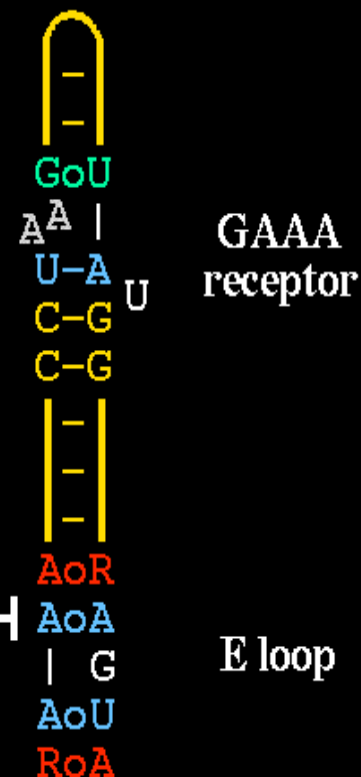
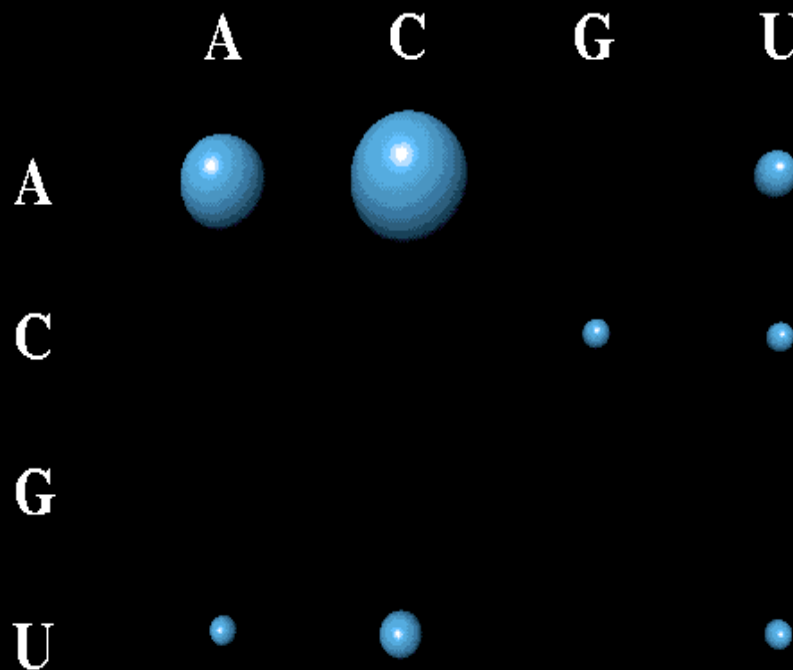
E loop 5'	GAAA receptor 5'	GAAA receptor 3'	E loop 3'
CAAUGAAAU-AU-CCUAAGU [6]		AUAUGGAUAGAGUAAUUU	
CGGGGAAAG-CAUCCUACAG [9]		CCAUGGUGCAAUAACCCU	
CGGGGAAAG-AA-CCUACGU [9]		AUAUGGUUCGAUUACCCU	
AGGGAAAACCA-CCUAAGU [6]		AUAUGGUC-GAGUAUCCU	

RAAA

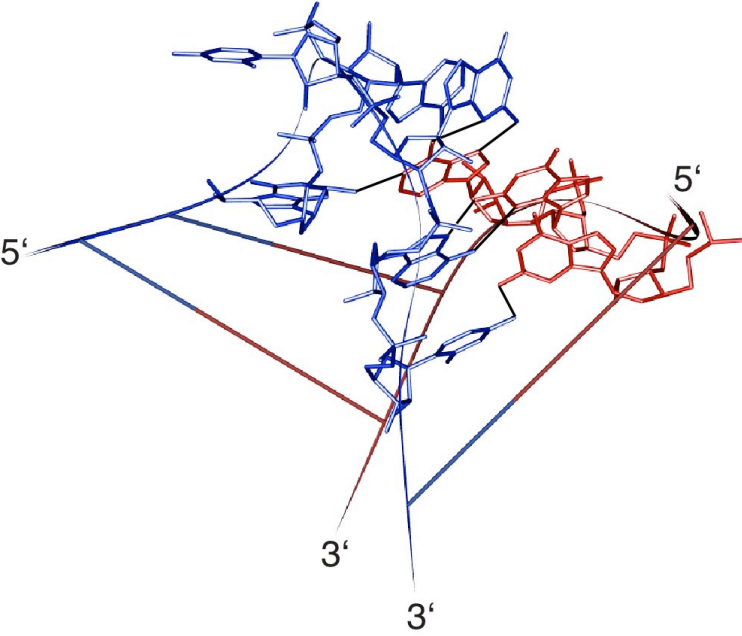
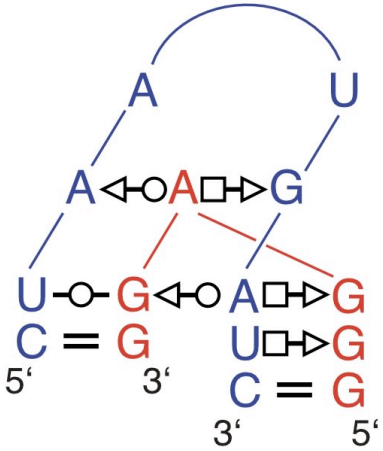
CCUAAG

UAUGG

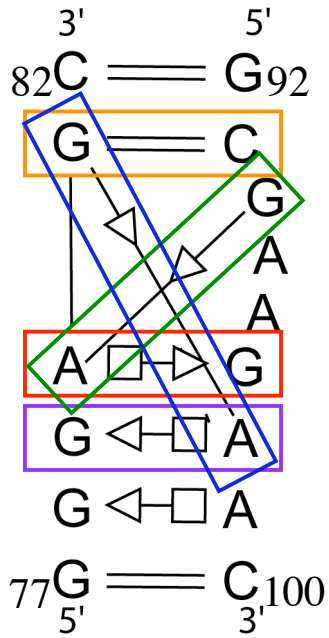
RAGUA



K-Turn



The K-turn motif



Watson-Crick

	cis	A	C	G	U
A	I4	i2	I3	I1	
C	I2	I6	I1	I5	
G	I3	I1		i2	
U	I1	I5	I2	I6	

Sugar-edge

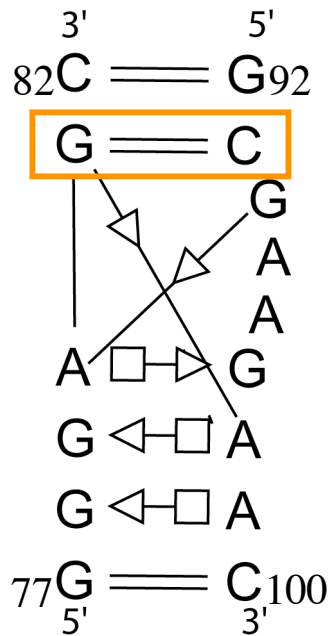
	trans	A	C	G	U
A	I1		(I2)		
C	I1		I2		
G	I1		I2		
U	I1		I2		

Hoogsteen

	trans	A	C	G	U
A	I1	I1	I1	I1	
C	I1	I1			I1
G				I2	
U	I2			I2	

Cis Watson-Crick/Watson-crick

The K-turn motif



Watson-Crick

	cis	A	C	G	U
A	I4	i2	I3	I1	
C	I2	I6	I1	I5	
G	I3	I1		i2	
U	I1	I5	I2	I6	

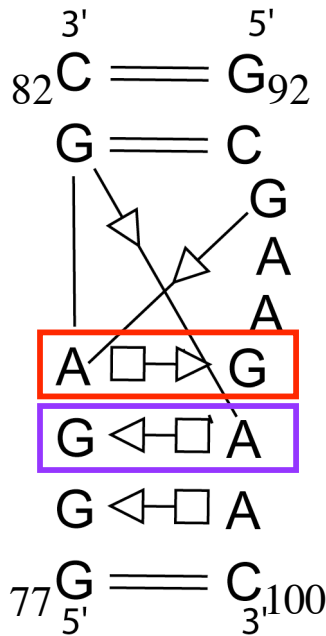
G85

trans	A	C	G	U	-
A			0 (7)	7 (0)	
C	0 (6)		801 (786)	0 (7)	0 (3)
G					
U	6 (0)		11 (17)		
-			0 (3)		3 (0)

C97

Trans Hoogsteen/Sugar-edge

The K-turn motif



Sugar-edge

trans	A	C	G	U
A	I1	I1	I1	I1
C	I1	I1		I1
G			I2	
U	I2		I2	

Hoogsteen

G97

trans	A	C	G	U	-
A	4 (4)		675 (673)	145 (145)	0 (2)
C					
G					
U					
-			0 (2)		2 (0)

A80

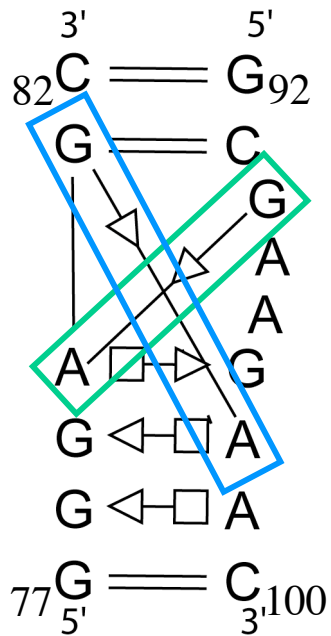
G79

trans	A	C	G	U	-
A	126 (125)	0 (1)	583 (580)	113 (112)	0 (3)
C					
G					
U			1 (1)		
-	0 (1)	1 (0)	0 (3)	0 (1)	3 (0)

A98

Cis Sugar-edge/Sugar-edge

The K-turn motif



Sugar-edge

trans	A	C	G	U
A	I1		(I2)	
C	I1		I2	
G	I1		I2	
U	I1		I2	

A80

G94

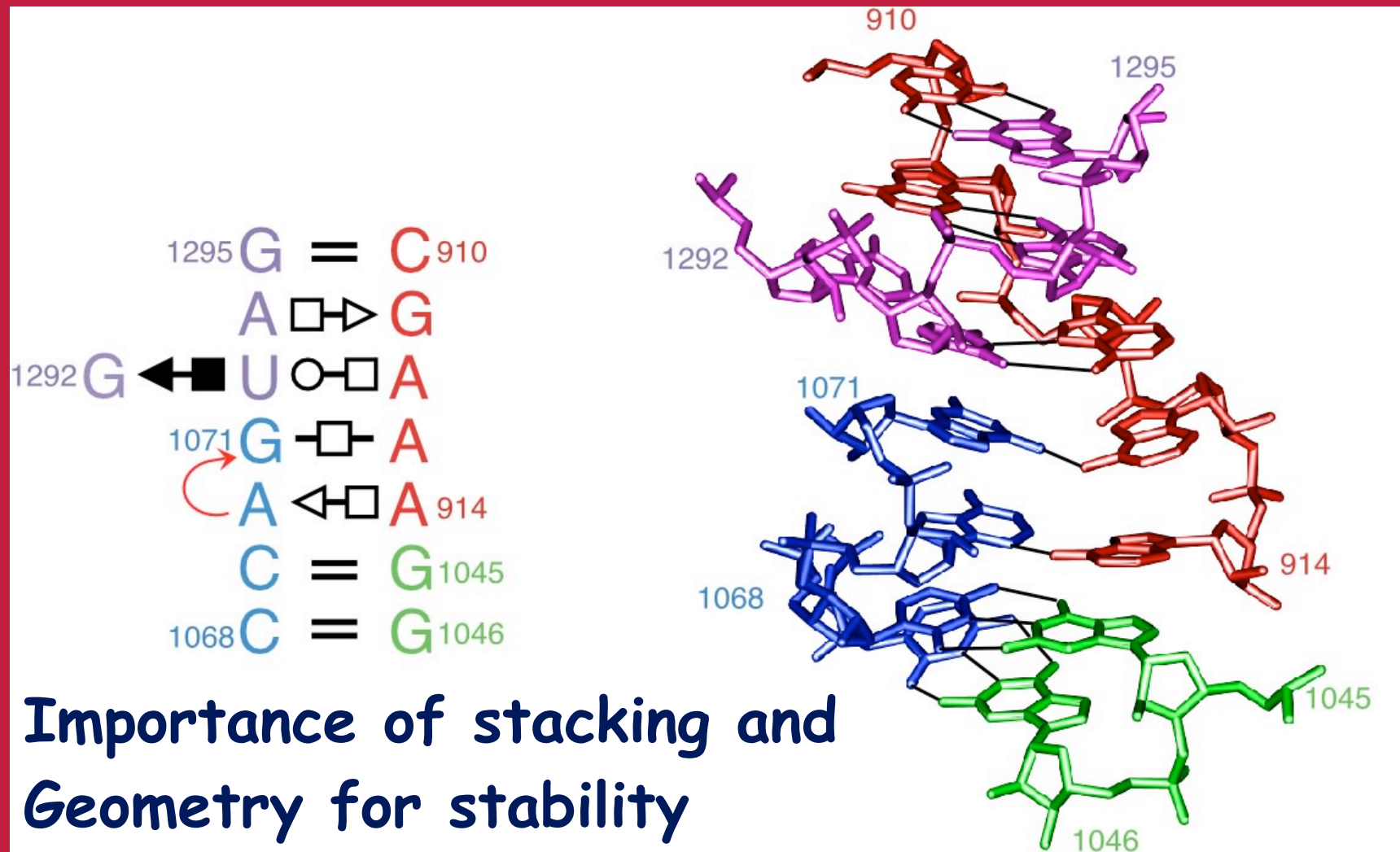
trans	A	C	G	U	-
A	38 (38)				
C	15 (15)				
G	130 (130)				
U	1 (1)				
-	640 (641)				3 (2)

A98

G81

trans	A	C	G	U	-
A	6 (6)				
C					
G	810 (807)			1 (1)	1 (4)
U	7 (7)				
-	0 (3)				3 (0)

Composite motifs : association of several strands



Importance of stacking and Geometry for stability

Structural Bioinformatics



Aligned Sequences
Organized according
To
Phylogeny

Corresponding
2D Structure
Drawings

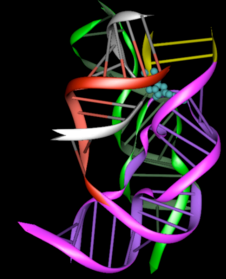
High Resolution
3D Structures

EVOLUTION

Isostericity matrices

Library of motifs

Levinthal's Paradox



$L = 100$ nucleotides $> 4^{100}$ (10^{60}) sequences

$1,8^{400}$ (10^{102}) structures 2D

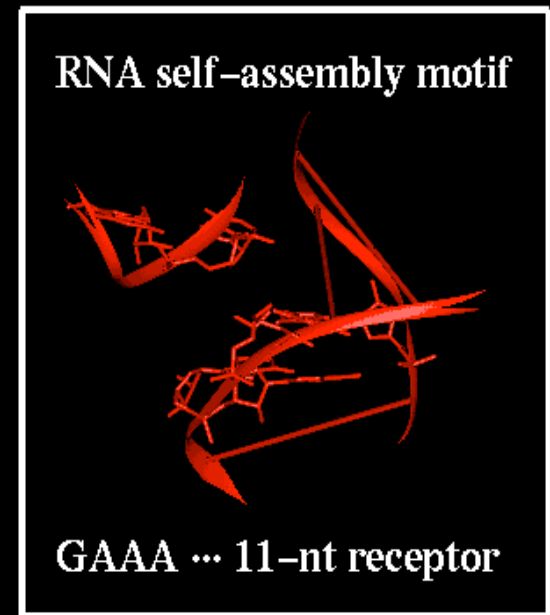
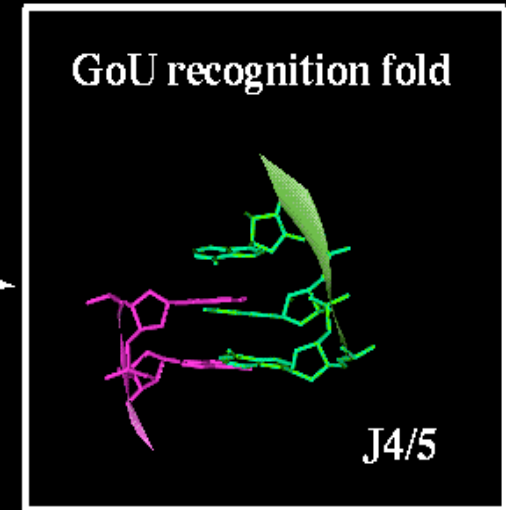
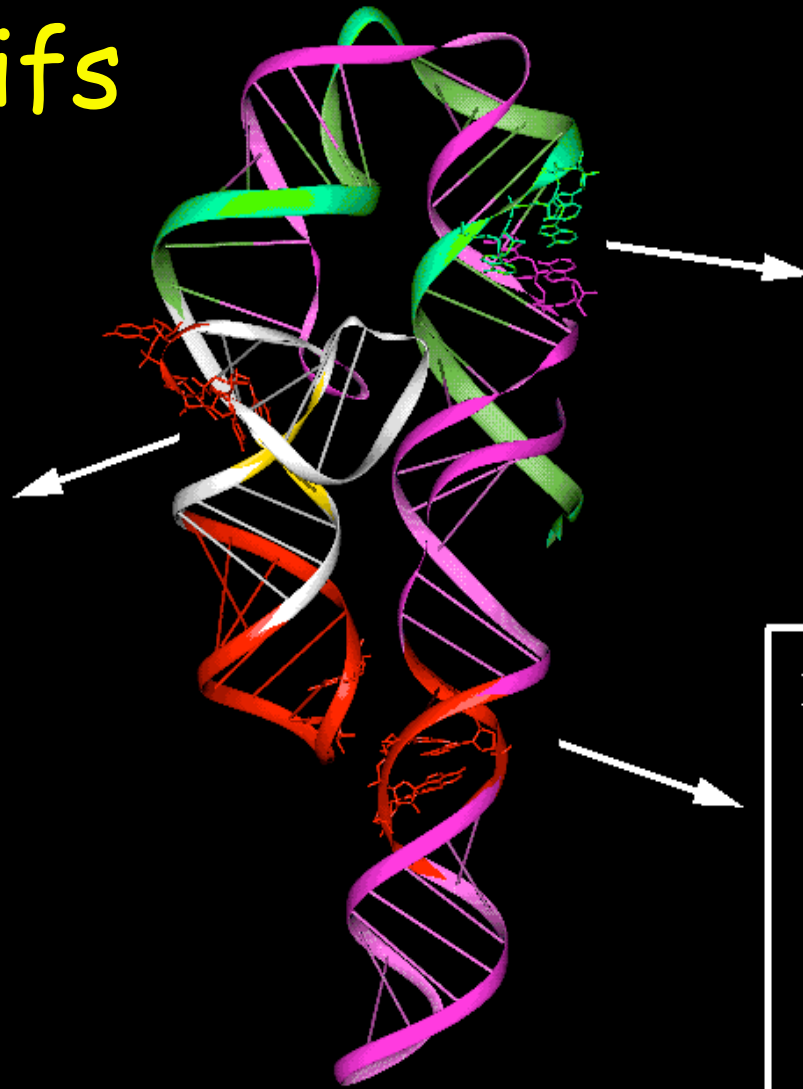
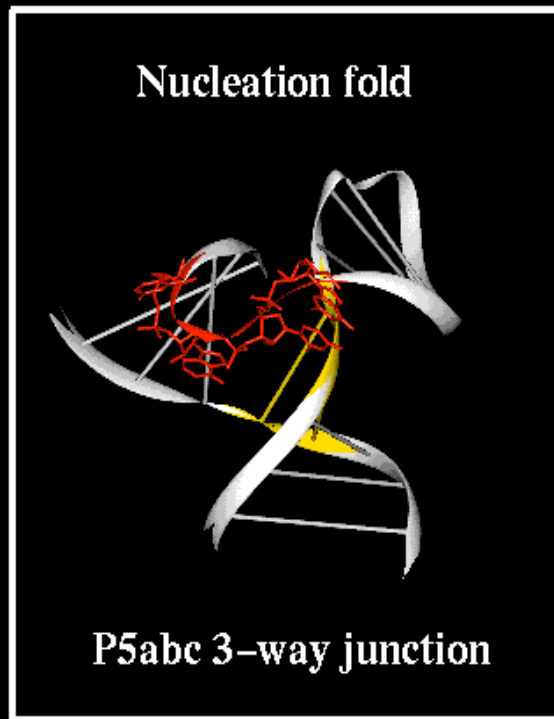
7 torsion angles / nucleotide

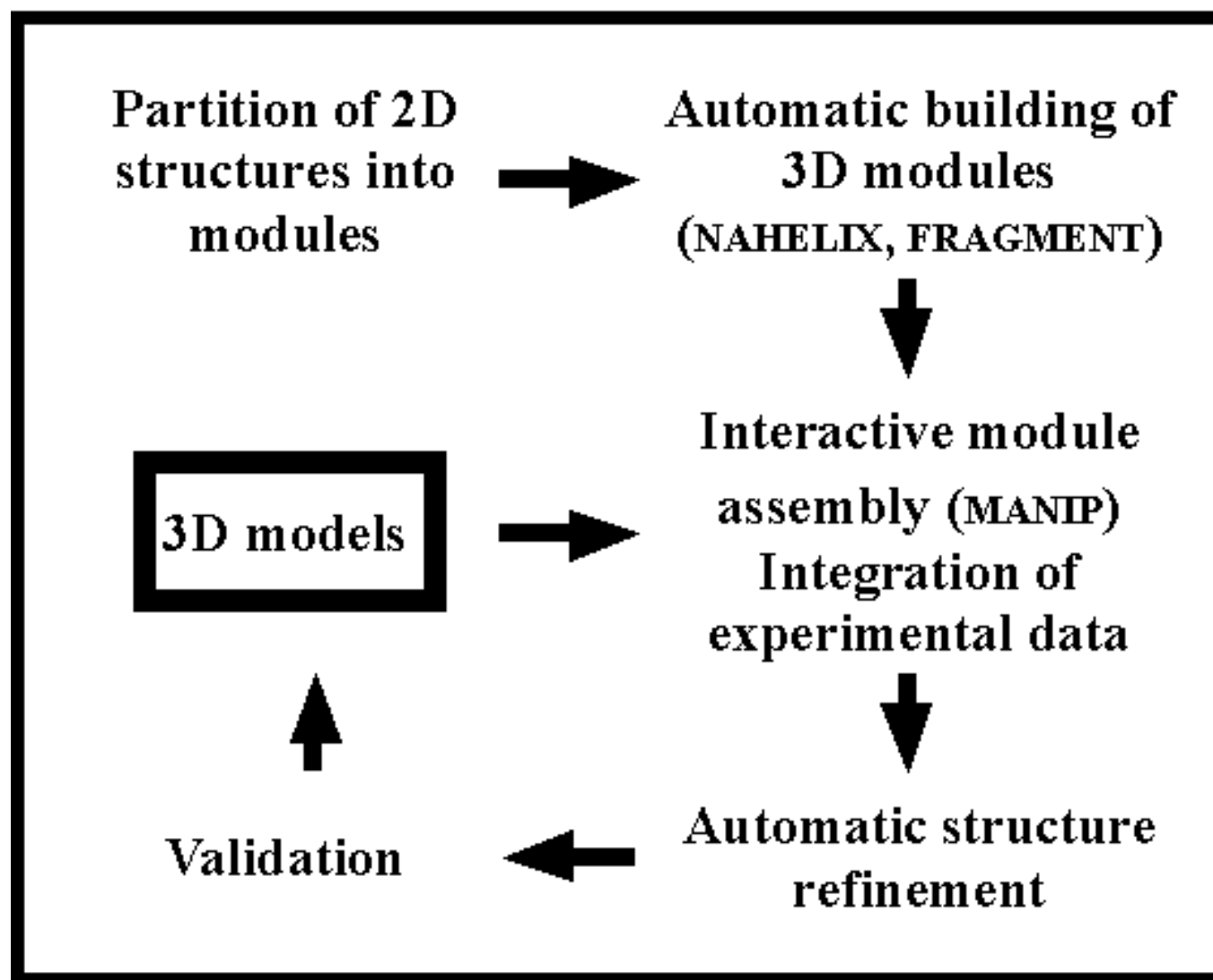
Torsion angles restricted & in the preferred conformers all the short range interactions are favourable

RNA Tectonics

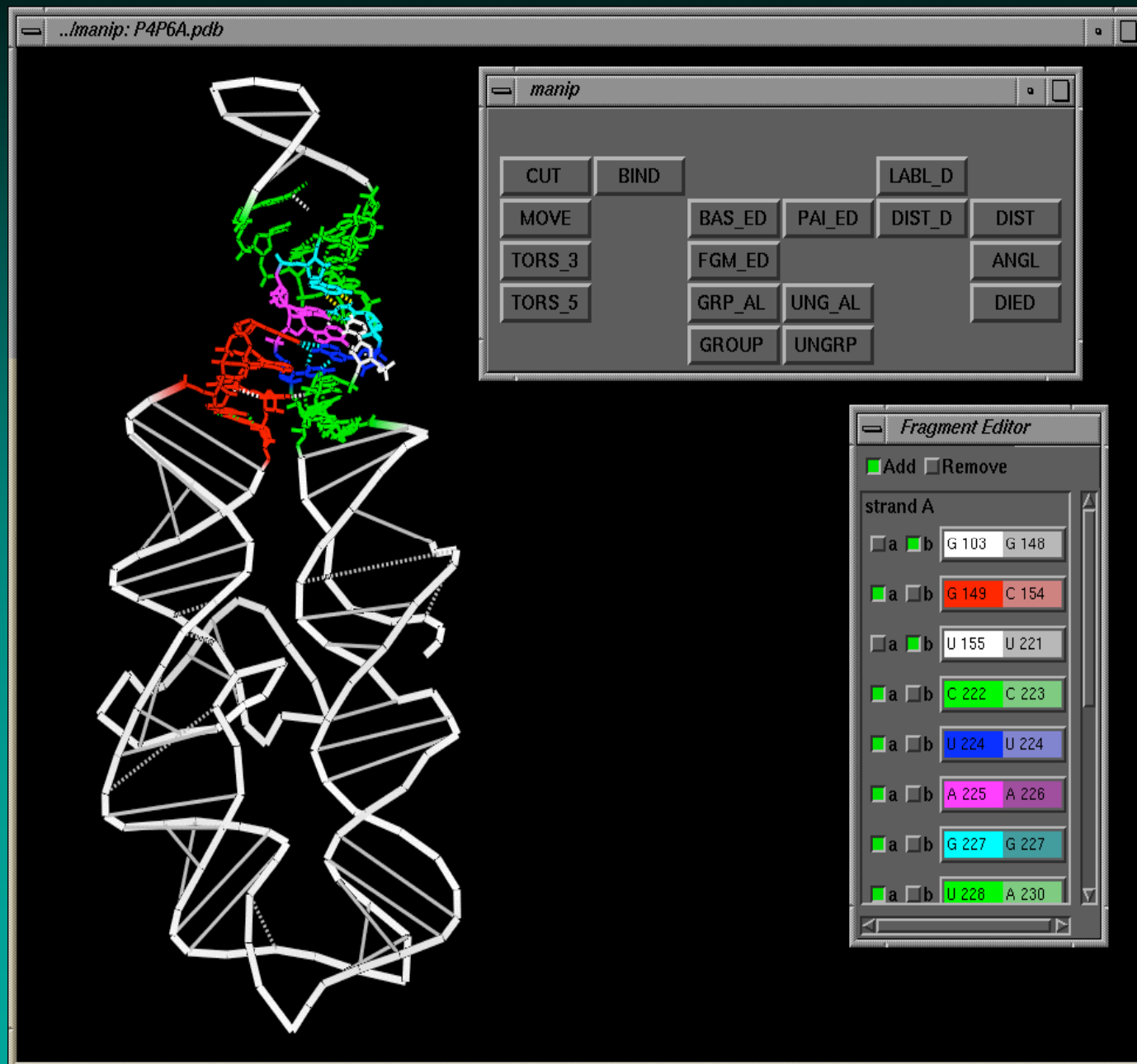
- **Modular and Hierarchical assembly of RNA molecules using motifs**

Each structure can be parsed in key motifs





Modelling tool : MANIP



NAHELIX

The screenshot displays the NAHELIX software interface. The main window, titled `..lmanip: CP3.hd`, shows a 3D molecular model of a nucleic acid structure. The structure is composed of several strands represented by colored sticks: red, blue, green, and cyan. Labels for specific atoms are visible: `O3* U153 A`, `P A204 C`, `O3* G163 B`, and `O3* U209 C`. A dialog box titled `NAHELIX` is open in the foreground, allowing for the configuration of the helix type and parameters.

NAHELIX Configuration Dialog:

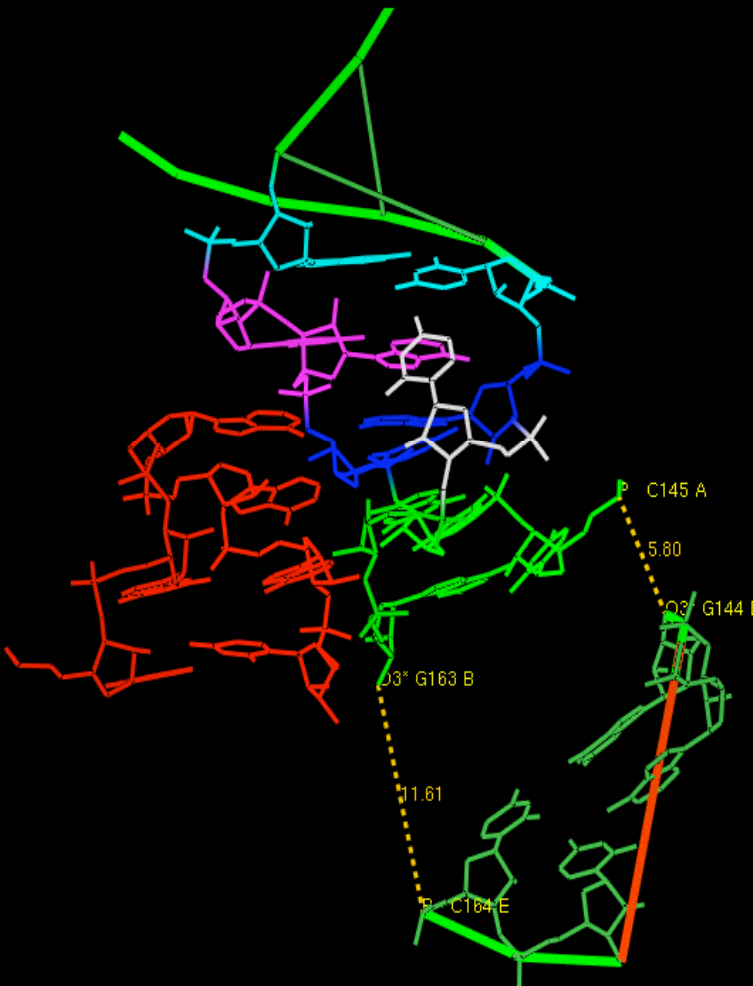
- Type of Helix:**
 - A-DNA
 - Z-DNA
 - triple-DNA
 - dA-dT
 - B-DNA
 - A-RNA
 - triple-RNA
 - dAT
- Number of residues in each strand:** 2
- Starting residue number in strand W:** 143
- Sequence of strand W:** AG
- Canonical Watson-Crick base pairing
- Starting residue number in strand C:** 164
- Output file:** (empty text field)
- Buttons:** OK, CANCEL

FRAGMENT

NUCLIN

NUCLSQ

..lmanip: CP4.hd



The image shows a 3D molecular model of a nucleic acid structure, likely RNA, with several strands and highlighted regions. The strands are color-coded: strand A (green), strand B (blue), strand C (orange), strand D (red), and strand E (purple). The model is displayed in a software window titled "Fragment Editor". The "Bases" panel on the right lists the sequence of bases for each strand, and the "Fragment Editor" panel shows the selected bases for each strand, with checkboxes for "Add" and "Remove".

Bases

- U 153
- strand B
- U 154
- C 155
- G 156
- G 157
- A 158
- U 159
- A 160
- U 161
- G 162
- G 163
- strand C
- A 204
- G 205
- A 206
- A 207
- A 208
- U 209
- strand D
- A 143
- G 144
- strand E
- C 164
- U 165

Fragment Editor

Add Remove

strand A

- a b C 145 C 146
- a b U 147 U 147
- a b A 148 C 149
- a b G 150 C 150
- a b U 151 U 153

strand B

- a b U 154 A 158
- a b U 159 U 159

strand C

- a b A 160 A 160
- a b U 161 U 161
- a b G 162 G 163

strand D

- a b A 204 U 209

strand E

- a b C 164 U 165

Labels in the model:

- C145 A
- G144 D
- G163 B
- C164 E
- 5.80
- 11.61

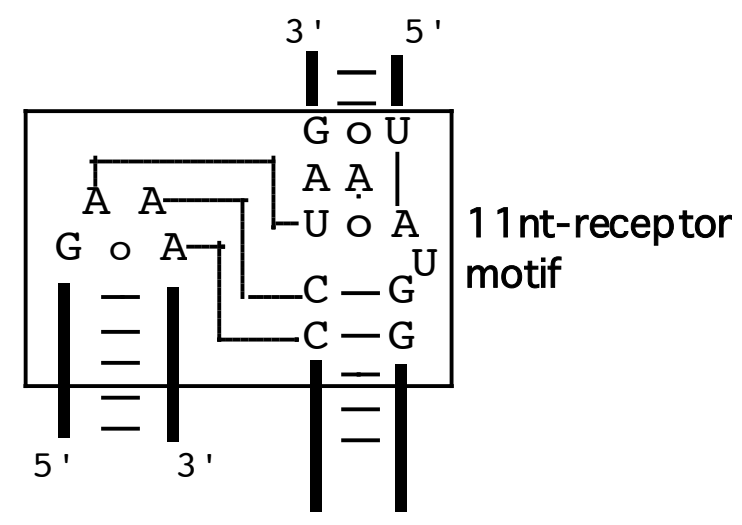
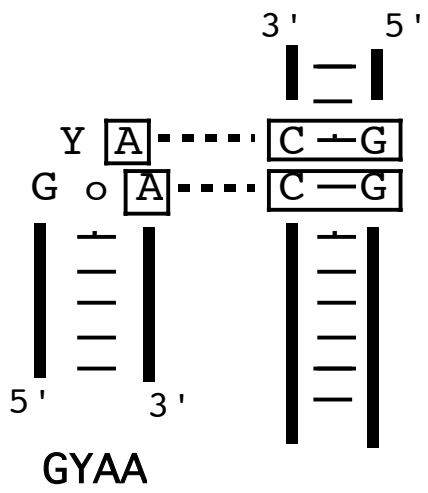
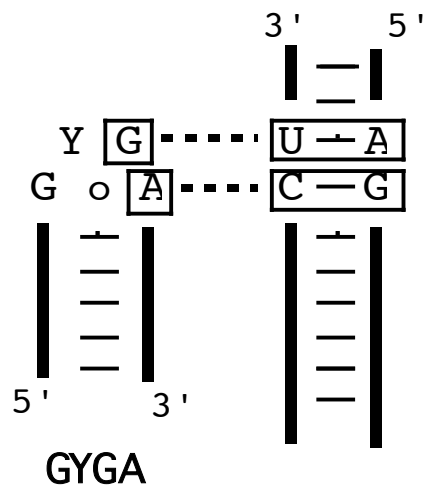
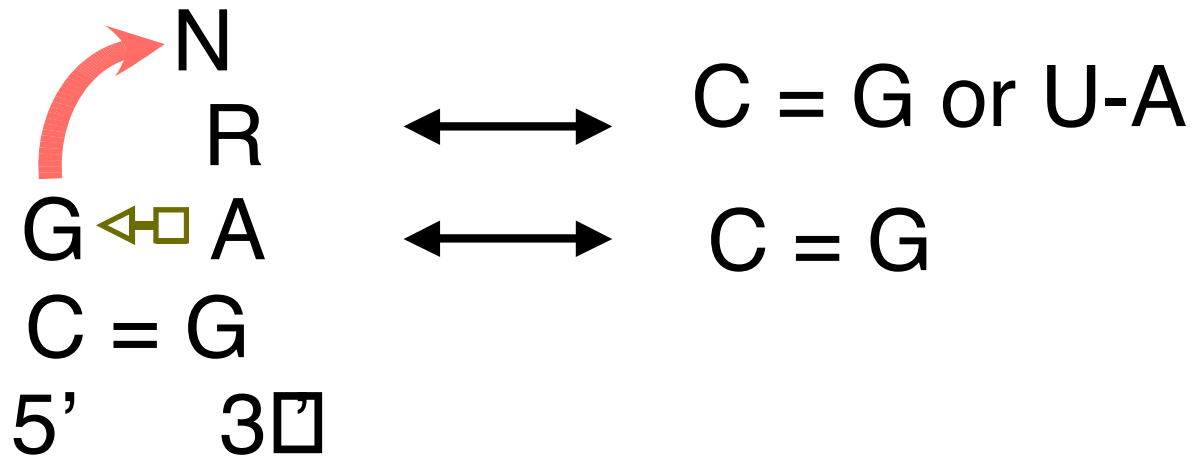
Properties of the modules:

- interfaces should be easily identified;
- they should interact via defined protocols;
- they should be easily modified and should evolve independently.

Properties of the Protocols:

- Rules of association between modules;
- Should be robust & should promote evolution.

GNRA - helix/receptors



Swap between GNRA/helix and loop-loop motifs

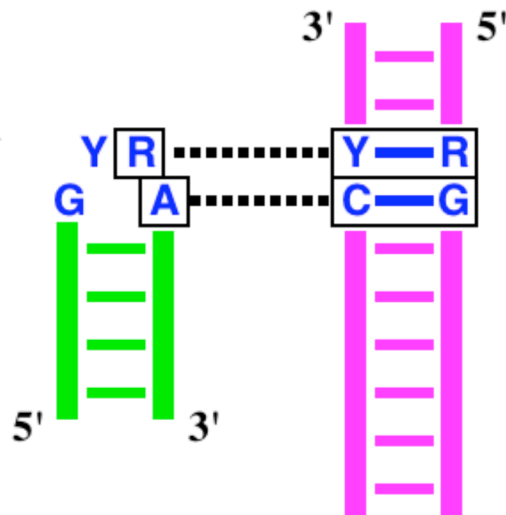
GYRA loop

AGAGUCGAUU [] GGGC-----GUGAGCCU [] GAUCGACUCU-----
 CAGGAUAGGG [] CGGG-----GCAACCCG [] CCCUAUCCUG-----

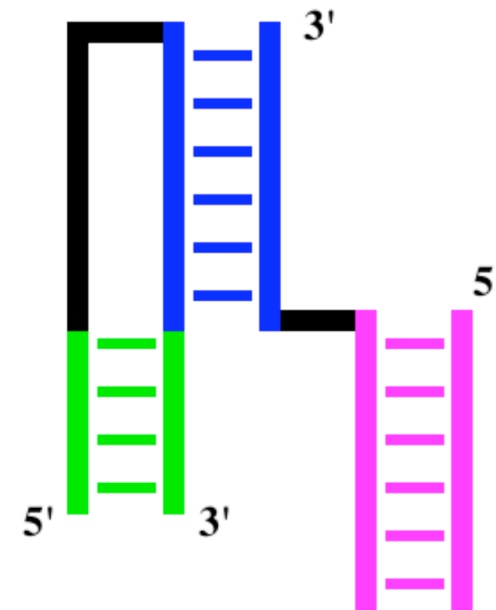
Pseudoknot

-----GCUGUC [] GAUG-UGAUAGCCAU [] GACAGCAUGCUAUC
 ---CGCUGUC [] GACGCUAAUAGACGUC [] GGCAGCGUUCUAUU

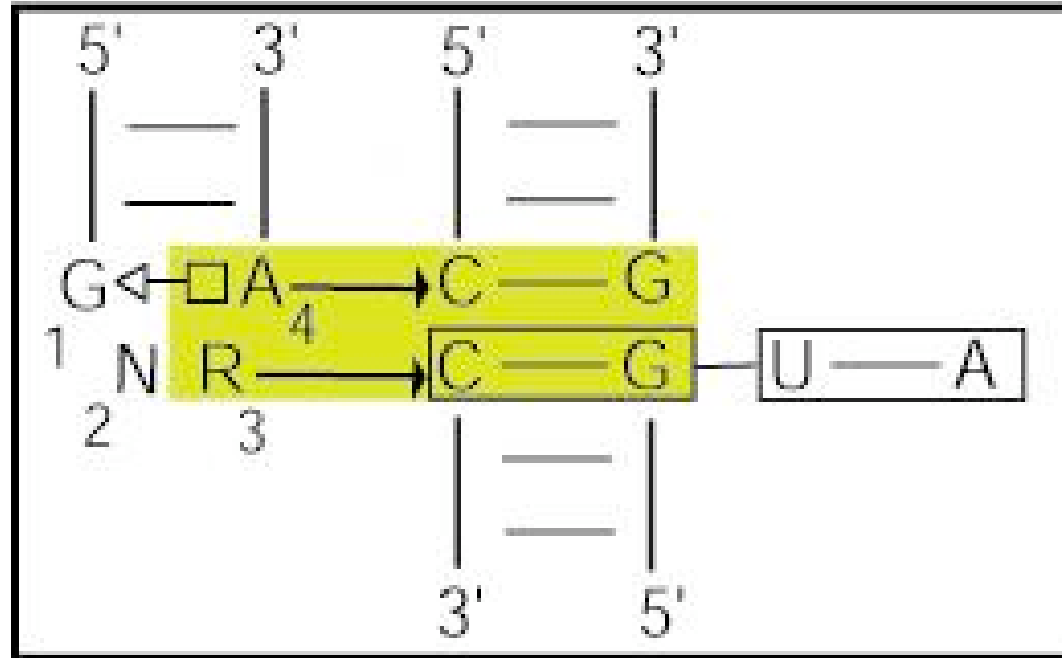
GYRA loop



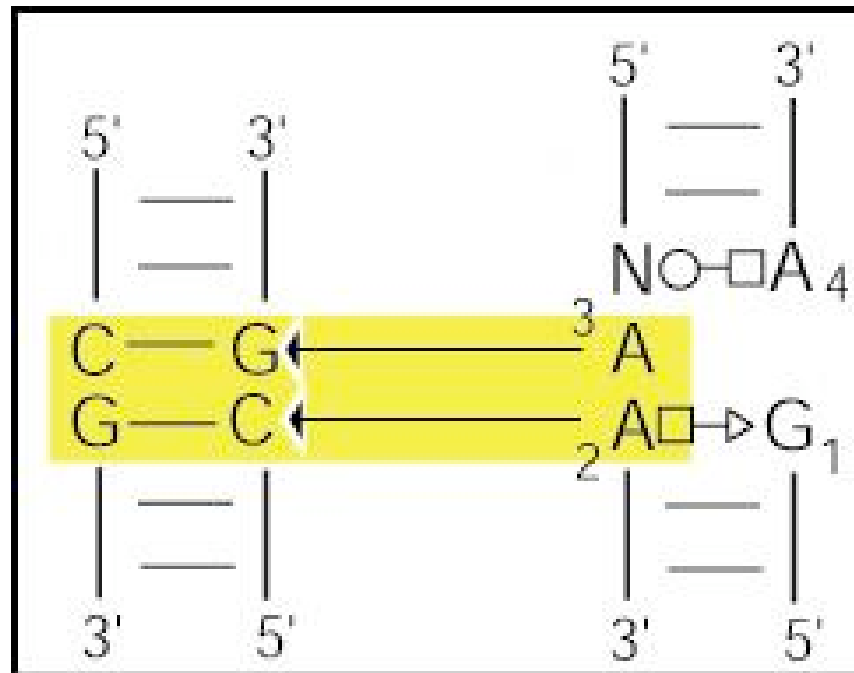
Pseudo knot



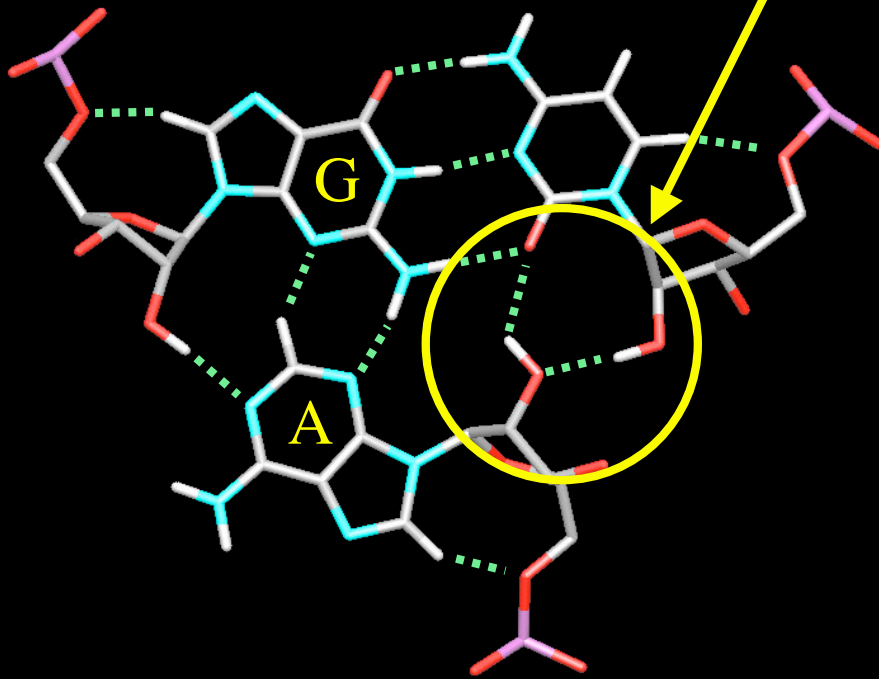
GNRA



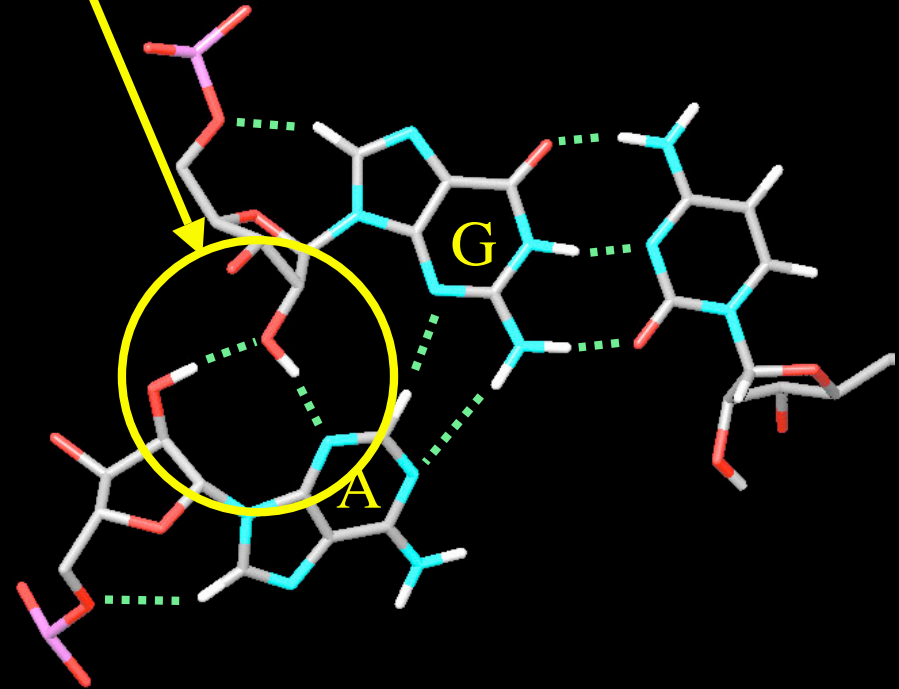
**A-rich
internal loop**



Sugar Edge/ Sugar Edge Ribose zipper



Trans and Parallel

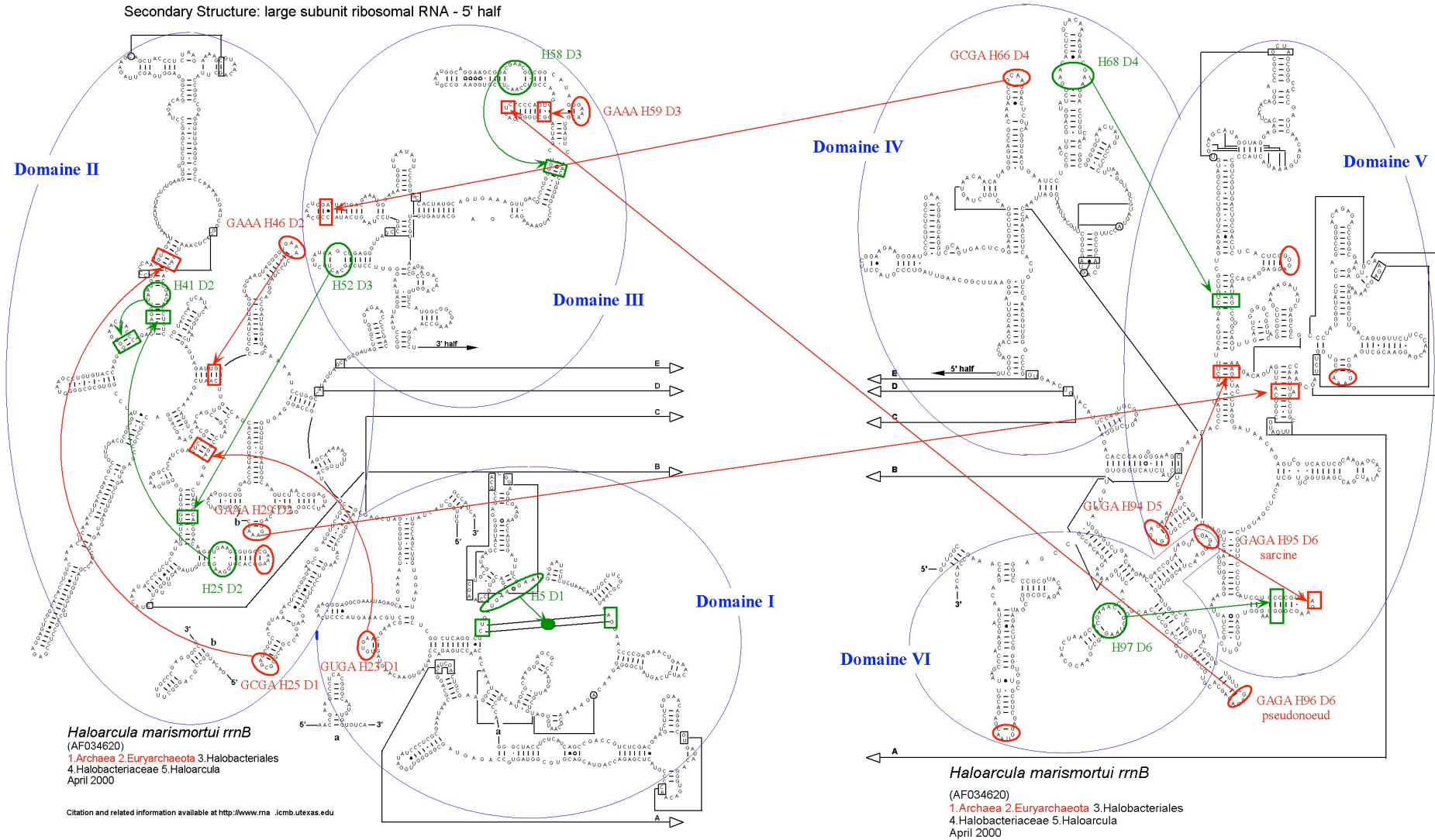


Cis and Anti-Parallel

Variety of A-minor motifs in 50S particle

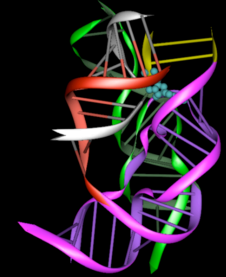
Secondary Structure: large subunit ribosomal RNA - 3' half

Secondary Structure: large subunit ribosomal RNA - 5' half



Citation and related information available at <http://www.ncbi.nlm.nih.gov/nucl/AF034620>

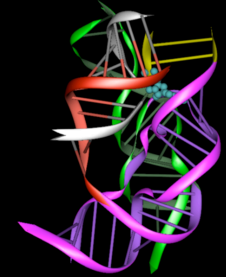
Laws of evolution



- ▶ Three-dimensional architectures evolve less with time than sequences
- ▶ Three-dimensional structures are dictated first by folding rules and secondarily by function

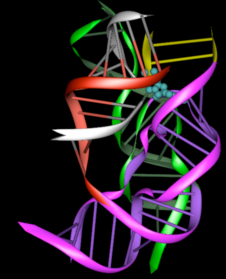
The phonetic structure of words are more stable than the meaning of words

Modelling algorithm



- 3D structure : assembly of fragments
- Stress 3D fold rather than sequence (inverse folding)
- Search for a «[consensus]» 3D fold (global architecture)

Modelling Constraints



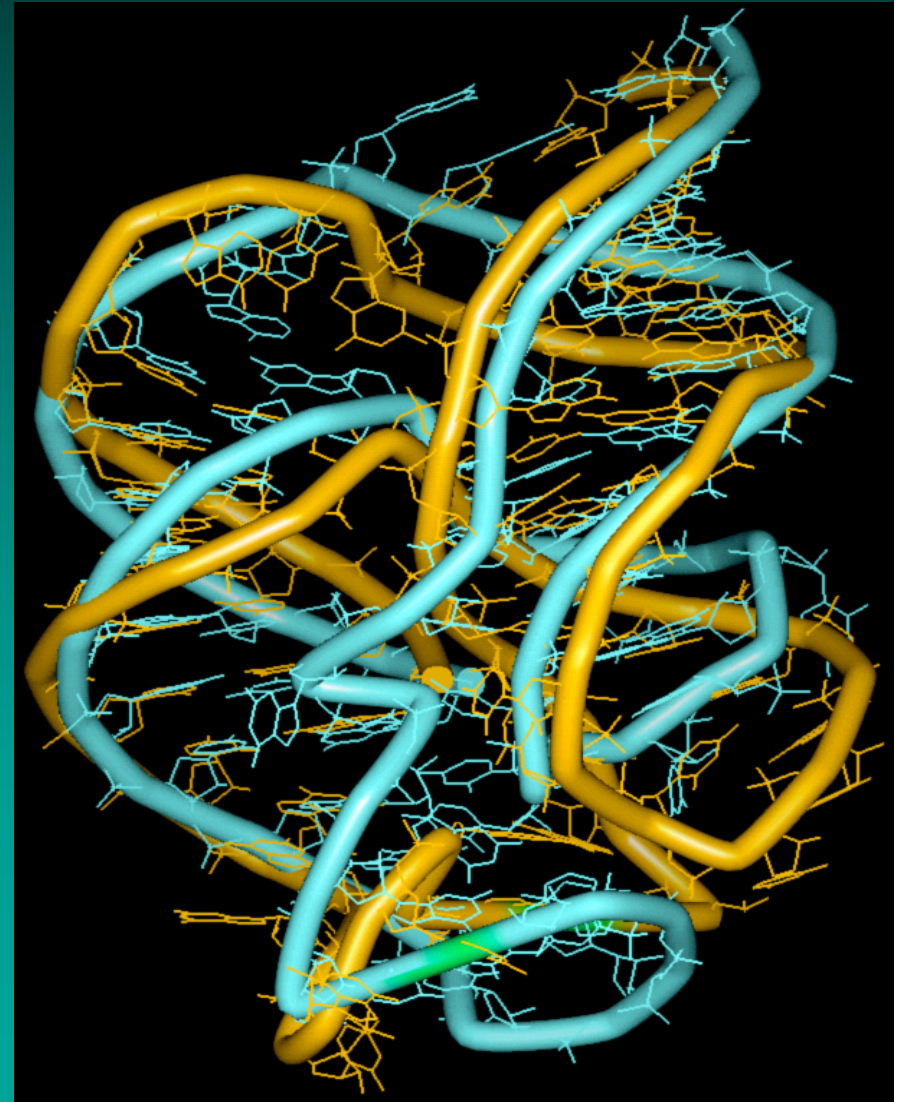
- 2D Topology (not strongly correlated with sequence) :
 - RNA is right-handed > right-handedness of stacks, of junctions
 - Type of junction : 3-way or 4-way...
 - Positions of single-strands w/r to the junction

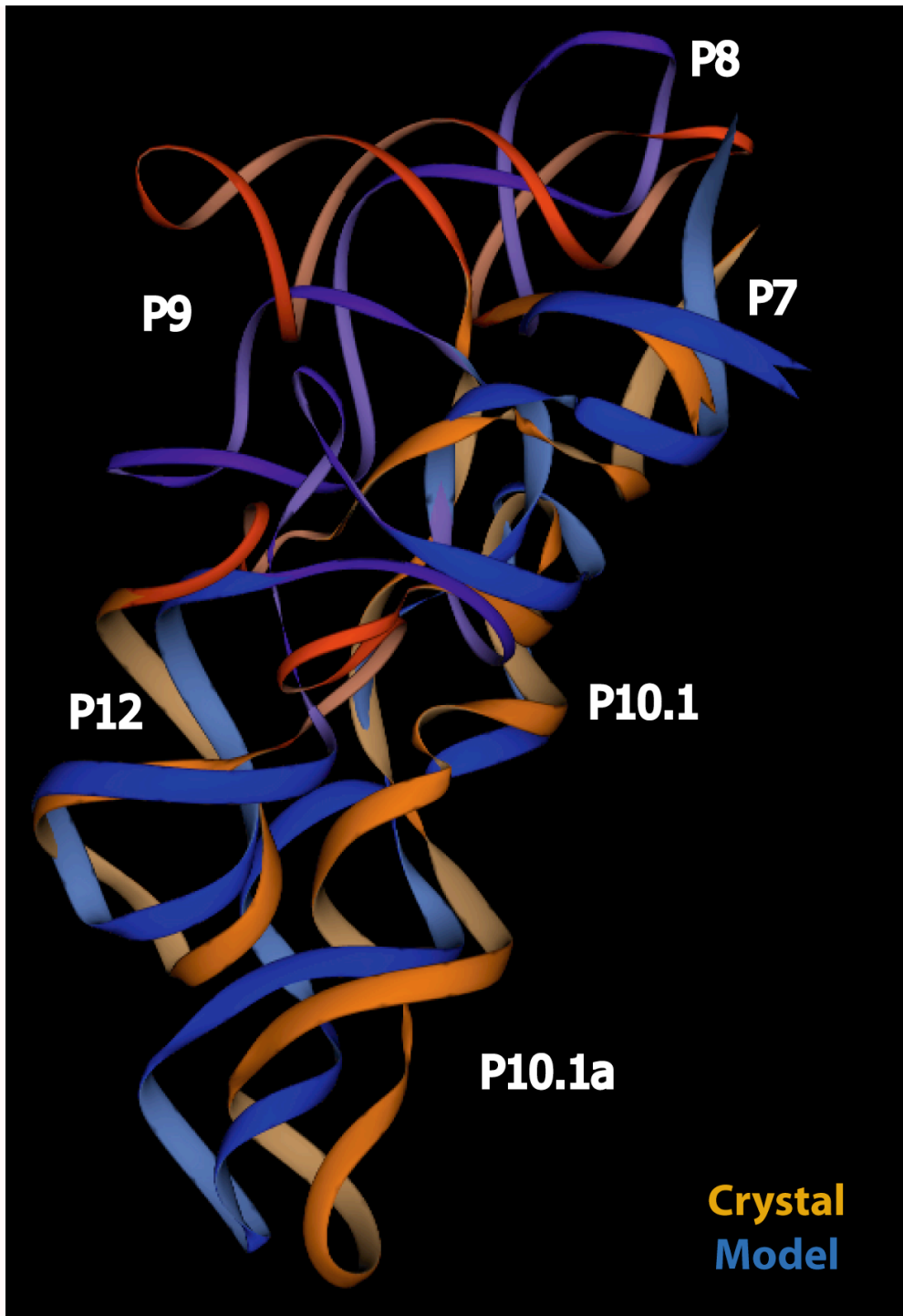
HDV ribozyme : X-ray vs. Model (r.m.s. 8.0 Å)

Ferré d'Amaré et al.
Nature 395, 567 (1998)

Tanner et al.
Current Biol. 4, 488 (1994)

**Two invariant G=C base pairs
were not modelled**





***rmsd* X-ray structure/model**

overall:	11.2 Å
minus P8-P9:	8.6 Å
minus P8-P9 L11/12	5.5 Å

***nrmsd* X-ray structure/model**

Carugo & Pongor (2001) *Prot Sci*

overall:	4.8 Å
minus P8-P9:	3.9 Å
minus P8-P9 L11/12	2.7 Å

**2003
1998**

Rms (> 1500 atoms)

9.06 Å

*Normalized rms
100 atoms)*

3.85 Å

